

Faculty and Staff Information

Chair – Santosh D'Mello, PhD	
Stephanie Aamodt	
Stephen Banks	15-17
Beverly Burden	
Matyas Buzgo	
Amy Erickson	
Mike Maguigan	
Elahe Mahdavian	
Stuart Nielsen	
Vonny Salim	
Peter Siska	
Stephanie Villalba	56-58

Santosh D'Mello, PhD



Professor & Chair PhD: University of Pittsburgh Postdoctoral training: Boston University Medical School; The Institute of Neurobiology, CNR – Rome Office: SC104A Tel: 318-795-2417 Fax: 318-797-5090 Email: santosh.dmello@lsus.edu

Teaching interests

Will be teaching *Neuroscience-I* (3 credits) and *Molecular basis of brain development* (3 credits) and degeneration starting Spring 2022 and Neuroscience-II in starting Fall 2022.

Research interests

Molecular mechanisms regulating neurodegeneration & brain development

Research in the lab is centered on understanding the molecular mechanisms regulating neurodegeneration. Specifically, primary cultures of neurons, transgenic and knockout mice, and animal models of neurological disease are used to study genes, proteins, and signal transduction pathways regulating neuronal cell death. We are also interested in identifying chemical compounds that protect the brain from neurodegeneration. The long-term objective of the laboratory's research is to develop strategies to prevent, treat, or cure degenerative diseases of the brain. Recently, we have expanded our interests to investigate neurodevelopmental disorders also. Our research has been funded by grants from the National Institutes of Health (NIH), Department of Defense, the National Science Foundation and private foundations. Our ongoing research on neurodegeneration and neurodevelopmental disorders is described below.

Neurodegenerative disorders

Neurodegenerative diseases such as Alzheimer's disease, Parkinson's disease (PD), and Huntington's disease (HD) are progressive and fatal disorders affect millions of individuals in the

U.S. alone costing the economy over \$100 billion annually. While there are drugs that can reduce the symptoms associated with some of these diseases (for example, Parkinson's disease), they do not slow down the relentless loss of neurons and therefore the disease progresses. Our lab is interested in identifying molecules that regulate the survival or death of neurons and whose altered function contributes to neurodegenerative disorders. Once identified, such molecules can then be targeted in the development of effective therapeutic strategies for these disorders. Much of our focus has been on histone deacetylases (HDACs) a family of 18 proteins initially identified based on their ability to repress gene expression through the deacetylation of histones, but which are now known to have a variety of other functions mediated through the deacetylation of non-histone proteins residing in the nucleus, cytoplasm or mitochondria. In studies supported by the NIH, we discovered that activation of one of the members of this family of proteins, HDAC3, plays a central role in promoting neurodegeneration. We are studying the mechanism by which HDAC3 promotes neurodegeneration.

The lab is also interested in FoxG1, a protein belonging to the Forkhead family of transcription factors that is critical for proper brain development where it controls the production of neurons by regulating proliferation of neural progenitor cells. Mice that lack FoxG1 have a severely underdeveloped brain and die early during gestation. But FoxG1 is highly expressed in the adult brain where its function had not been studied. We recently found that FoxG1 maintains the survival of mature neurons. We have been investigating the molecular mechanism through which the activity of FoxG1 is regulated and the mechanism by which FoxG1 affects other molecules to maintain the survival of neurons. As part of an NIH-funded project, we generated transgenic mice that express elevated levels of FoxG1. These mice will be used to test whether elevated FoxG1 can protect mice against neurodegenerative diseases such as Huntington's disease. Another Forkhead protein of interest to the lab is FoxP1. FoxP1 is expressed selectively in the striatum and cortex, two regions of the brain that are selectively degenerate in HD. In studies funded by the NIH we found that FoxP1 expression is reduced in the striatum of HD patients and in HD mouse models. This reduction likely contributes the the loss of neurons in HD. In humans, FoxP1 mutations cause mental retardation and other cognitive deficits.

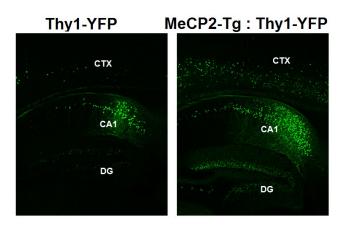
In addition to understanding the molecular biology of neurodegeneration the lab has been identifying chemical compounds that protect neurons from death. This drug discovery effort has led to the identification of several indolone and benzoxazine compounds that are highly protective in cell culture models and animal models of neurodegenerative diseases. Exactly how these neuroprotective compounds act is an area of interest.

Neurodevelopmental disorders

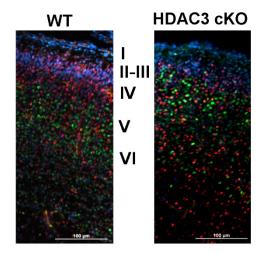
We have recently become interested in MeCP2, a gene that can repress gene transcription globally as well as locally. Loss-of-function mutations in the MeCP2 gene cause Rett syndrome, a neurodevelopmental disorder characterized by a slowing of development, loss of purposeful use of the hands, distinctive hand movements, slowed brain and head growth, problems with walking, seizures, and intellectual disability. On the other hand, elevated activity of MeCP2 as a result of gene duplication or triplication causes another neurological disorder called MeCP2 duplication syndrome. Patients with this disorder are born normal but then display progressive mental retardation, spasticity, epilepsy, and die at adulthood. We are studying MeCP2

duplication syndrome using transgenic mice that make 3-4 times more MeCP2 than normal. Like patients with MeCP2 duplication syndrome, these mice display neurological deficits and die early in adulthood. The mice display neuronal loss in certain brain regions coincident with the neurological symptoms and just before they die. We are characterizing other abnormalities in the MeCP2 transgenic brain with the goal of getting a better understanding of why human patients with MeCP2 duplication syndrome suffer the neurological phenotype that they do. A recent discovery we have made is that astrocytes within certain brain regions of the MeCP2 transgenic mice have high levels of a protein called GFAP. Interestingly, increased GFAP production is the primary cause of another neurological brain disorder called Alexander disease, characterized by spasticity, mental retardation, and seizures. These symptoms are also observed in many patients with MeCP2 duplication syndrome. We are exploring whether MeCP2 duplication syndrome and Alexander disease share mechanistic commonalities.

While abnormal function of HDAC3 contributes to neurodegeneration, recent research in the lab on brain-specific conditional knockout mice has revealed that HDAC3 plays an essential role in brain development. Neuronal migration and formation of proper lamination in the cortex is disrupted in mice lacking HDAC3 in the brain. Exactly how HDAC3 regulates proper brain development is being studied.



Supernumerary neurons in the hippocampus of MeCP2 transgenic mice



Disrupted lamination in the cortex of HDAC3 conditional knockout mice

Selected publications

Wang L, Ankati H, Akubathini S, Balderamos M, Storey C, Patel AV, Kretzschmar D, Biehl ER, D'Mello SR. (2010) 1, 4- benzoxazine compounds as novel neuroprotective agents. *J. Neurosci. Res.* 88: 1970-1984.

Ankanti H, Akubathini SK, D'Mello SR., Biehl ER (2010) Synthesis of 2-Benzylidene and 2-Hetarylmethyl Derivatives of 2H-1,4-Benzoxazin-3-(4H)-ones as Neuroprotective Agents" *Synth. Communications* 40: 2364–2376 Zhao K, Ippolito G, Wang L, Price V, Kim MH, Cornwell G, Fulenchek S, Breen GA, Goux WJ, D'Mello SR. (2010) Neuron-selective toxicity of tau peptide in a cell culture model of neurodegenerative tauopathy: essential role for aggregation in neurotoxicity. *J Neurosci Res.* 88:3399-3413.

Chen HL, D'Mello SR. (2010) Induction of neuronal cell death by paraneoplastic Ma1 antigen. J Neurosci Res. 88:3508-3519.

Dastidar SG, Landrieu PM, D'Mello SR. (2011) FoxG1 Promotes the Survival of Postmitotic Neurons. *J Neurosci*. 31:402-413.

Ma C, D'Mello SR. (2011) Neuroprotection by histone deacetylase-7 (HDAC7) occurs by inhibition of c-jun expression through a deacetylase-independent mechanism. *J. Biol. Chem*. 286:4819-4828.

Bardai FH, D'Mello SR. (2011) Selective toxicity by HDAC3 in neurons: Regulation by Akt and GSK3^[2]. *J. Neurosci.* 31:1746-51. *In the list of 50 most downloaded Journal of Neuroscience papers, March 2011*

Garcia-Oscos F, Salgado H, Hall S, Thomas F, Farmer GE, Bermeo J, Galindo LC, Ramirez RD, D'Mello S, Rose-John S, Atzori M. (2011) The stress-induced cytokine interleukin-6 decreases the inhibition/excitation ratio in the rat temporal cortex via trans-signaling. *Biol Psychiatry.* 71:574-582.

Ghosh Dastidar S, Bardai F, Ma C, Price V, Rawat V, Verma P, Narayanan V, D'Mello SR. (2012) Isoform-specific toxicity of Mecp2 in postmitotic neurons: Suppression of neurotoxicity by FoxG1. *J. Neurosci.* 32:2846-2855.

Ghosh Dastidar S, Narayanan S, Stifani S, D'Mello SR. (2012) Transducin-like enhancer of Split-1 (TLE1) combines with Forkhead box protein G1 (FoxG1) to promote neuronal survival. J Biol Chem. 287:14749-14759.

Bardai FH, Price V, Zaayman M, Wang L, D'Mello SR. (2012) Histone deacetylase (HDAC1) is a molecular switch between neuronal survival and death. *J Biol Chem.* 287:35444-35453. *Paper of the week (Oct 2012), Best neuroscience paper (2012)*

Price V, Wang L, D'Mello SR (2013). Conditional deletion of HDAC4 in the CNS has no major effect on brain architecture or neuronal viability. *J. Neurosci. Res.* 91:407-15.

Bardai FH, Verma P, Smith C, Rawat V, Wang L, D'Mello SR. (2013) Disassociation of HDAC3 from normal huntingtin underlies mutant huntingtin neurotoxicity. *J. Neurosci.* 33:11833-11838.

Rangaswamy S, D'Mello SR, Narayanan V (2013) Epigenetics, Autism Spectrum, and Neurodevelopmental Disorders. *Neurotherapeutics* 10:742-756.

Verma P, Pfister, JA, Mallick S, D'Mello SR (2014) HSF1 protects neurons through a novel trimerization and HSP independent mechanism. *J Neurosci.* 34:1599-1612.

Mallick S. and D'Mello SR. (2014) JAZ (Znf346), a SIRT1-interacting protein protects neurons by stimulating p21 (WAF/CIP1) expression. *J. Biol. Chem.* 289:35409-35420.

Norwood J, Franklin JM, Sharma D, D'Mello SR. (2014) Histone deacetylase-3 is necessary for proper brain development. *J. Biol. Chem.289*: 34569-34582.

Garcia-Oscos F, Peña D, Housini M, Cheng D, Lopez D, Borland MS, Salgado-Delgado R, Salgado H, D'Mello S, Kilgard MP, Rose-John S, Atzori M. (2015) Vagal nerve stimulation blocks interleukin 6-dependent synaptic hyperexcitability induced by lipopolysaccharide-induced acute stress in the rodent prefrontal cortex. *Brain Behav Immun.* 43: 149-158.

Sharma D, Kim MS, D'Mello SR (2015). Transcriptome profiling of expression changes during neuronal death by RNA-Seq. *Exp. Biol. Med.*. 240:242-51. *Selected as the feature article for the February 2015 issue. Was also featured in a press release.*

Rawat V, Goux W, Piechaczyk M and D'Mello (2016) C-Fos protects neurons through a noncanonical mechanism involving HDAC3 interaction: Identification of a 21 amino acid fragment with neuroprotective activity. *Mol. Neurobiology* 53:1165-1180.

Pfister J and D'Mello SR. (2015) Insights into the regulation of neuronal viability by Nucleophosmin/B23. *Experimental Biology and Medicine* 240:774-786.

Smith C and D'Mello SR (2016) Cell and context-dependent effects of the heat shock protein DNAJB6 on neuronal survival. *Mol. Neurobiol.* 53: 5628-5639

Pfister JA and D'Mello SR (2016) Regulation of neuronal survival by nucleophosmin 1 (NPM1) is dependent on its expression level, subcellular localization and oligomerization status. *J. Biol. Chem*.291:20787-20797.

Louis Sam Titus ASC, Yusuff T, Cassar M, Thomas E, Kretzschmar D, D'Mello SR (2017) Reduced Expression of Foxp1 as a Contributing Factor in Huntington's Disease. *J. Neurosc*i.37:6575-6587.

Thomas EA and D'Mello SR (2018). Complex neuroprotective and neurotoxic effects of histone deacetylases. *J. Neurochem*.145:96-110.

Qu, Z and D'Mello SR. (2018) Proteomic analysis identifies NPTX1 and HIP1R as potential targets of HDAC3-mediated neurodegeneration. *Exp. Biol. Med.* 243:627-638

Montgomery KR, ASC Louis Sam Titus, Wang L, D'Mello SR (2018) Elevated MeCP2 in Mice Causes Neurodegeneration Involving Tau Dysregulation and Excitotoxicity: Implications for the Understanding and Treatment of MeCP2 Triplication Syndrome. *Molecular Neurobiology* 55:9057-9074.

Qu Z, Xuan Z, D'Mello SR. Neuroprotection by Heat Shock Factor-1 (HSF1) and Trimerization-Deficient Mutant Identifies Novel Alterations in Gene Expression (2018). *Sci Reports* 8(1):17255.

Pfister JA, Ma Chi, D'Mello SR (2019) Catalytic-independent neuroprotection by SIRT1 is mediated through interaction with HDAC1. *PLosOne* 14(4):e0215208

Louis Sam Titus ACS, Sharma D, D'Mello SR. (2020) The BDNF and Npas4 genes are targets of HDAC3-mediated transcriptional repression. *BMC Neuroscience* 28;20(1):65.

D'Mello SR. (2020) Regulation of CNS Development by Class I HDACs. *Developmental Neuroscience*. 41:149-165.

D'Mello SR.(2020) Histone deacetylase-3: Friend and foe of the brain. *Exp Biol Med* 245:1130-1141.

D'Mello SR and Kindy M. (2020) Overdosing on iron: Elevated iron and degenerative brain disorders. *Exp Biol Med* 245:1444-1473

D'Mello SR (2021) When Good Kinases go Rogue: GSK3, p38 MAPK and CDKs as Therapeutic Targets for Alzheimer's and Huntington's disease. International Journal of Medical Sciences 22(11):5911. doi: 10.3390/ijms22115911).

D'Mello SR. (2021) MECP2 and the Biology of MeCP2 Duplication Syndrome. *J. Neurochem*. doi: 10.1111/jnc.15331. (*Online ahead of print*).

Stephanie Aamodt, PhD



Professor Department of Biological Sciences Louisiana State University in Shreveport Phone: 318.797.5214 Fax: 318.797.5222 Office: SC 114a e-mail: <u>stephanie.aamodt@lsus.edu</u>

Professional Experience

Louisiana State University-Shreveport, Shreveport, LA 2005-present Professor, Dept. of Biological Sciences 2015-2020 Director, LSUS Quality Enhancement Plan 2000-2005 Chair, Department of Biological Sciences 1998-2005 Associate Professor, Dept. of Biological Sciences 1993-98 Assistant Professor, Dept. of Biological Sciences 1992-93 Instructor, Depts. of Biological Sciences and Chemistry 1991-92 Adjunct Instructor, Depts. of Biological Sciences and Chemistry

Louisiana State University Health Sciences Center-Shreveport, Shreveport, LA 2005-2006 Sabbatical, Dr. Tony Giordano, Director of the Biotechnology Program, mentor.

Tufts University School of Medicine, Boston, MA, 1977-79 Research Assistant, Dept. of Biochemistry and Pharmacology.

Education and Training

Postdoctoral Research Associate, Northwestern University School of Medicine, Dept. of Cell Biology and Anatomy, Chicago, IL, 1984-87

PhD in Molecular Biology, Vanderbilt University, Nashville, TN, 1984

BS in Biology, Bates College, Lewiston, ME, 1977

Publications

E. Aamodt and S. Aamodt (2006) Neural Specification and Differentiation in the Neurobiology of *C. elegans*, Academic Press, 74-99.

S. Aamodt, P. Polk and E. Aamodt (1999) Deletion Mutation of *C. elegans* PTL-1, a Tau Homologue. Molecular Biology of the Cell (abstract).

S. Aamodt (1997) PTL-1, the tau-like microtubule-associated protein from *C. elegans*. Dev. Biol. 186: 302 (abstract).

J.B. McDermott, S. Aamodt and E. Aamodt (1996) *ptl-1*, a *Caenorhabditis elegans* gene whose products are homologous to the tau microtubule associated proteins. Biochemistry 35 (29): 9415-9423.

Teaching Experience: Undergraduate

Courses recently taught:

Fundamentals of Biology I (BIOS 110, for majors) Summer 2014-2019 Fundamentals of Biology II (BIOS 120, for majors) Fall 1995-2004, 2005, 2006, 2007, 2009, 2011, 2014, 2019; Spring 2008, 2009, 2013, 2020, 2021 Summer 2001, 2004, 2008, 2020, 2021

Endocrinology (BIOS 480), Fall 2007, 2009, 2011, 2013, 2015, 2017, 2020

Histology, Summer 2018-2021 Histology Laboratory, Summer 2018, 2019

Immunology, (BIOS 321/421) Spring 2002, 2004, 2009, 2010, 2012, 2014-2021 Summer 2003, 2008, 2011, 2013, Fall 2006 Immunology Laboratory, Fall 2006, Spring 2009, 2010, 2012, 2014, 2019; Summer 2011, 2013

Introduction to Biological Inquiry (BIOS 210) Spring 2008-2016, Summer 2015-2017, Fall 2008-2016

Parasitology (BIOS 380), Fall 2008, 2010, 2012, 2014, 2017, 2019, 2021 Parasitology Laboratory, Fall 2008, 2012, 2014

Principles of Genetics (BIOS 363) Spring 2020, 2021, Fall 2021 Principles of Genetics Laboratory (BIOS 363L) Fall 1995, 2020, Spring 2020, 2021

Vertebrate Embryology/Developmental Biology (BIOS 375/475) Spring 1994, 1996-2000, 2002, 2004. 2008, 2013, 2016, 2017, 2022, Fall 2010 Vertebrate Embryology Laboratory (BIOS 375L/475L) Spring 1994, 1996-2000, 2002, 2008, 2013, 2022 Courses previously taught: Biochemistry (CHEM 383,384) Spring 1991-Fall 1992 Biochemistry (BCHM 410) Fall 1999 Biochemistry Laboratory (BCHM 410L) Fall 1999

Freshmen Seminar (FS 104), Fall 2009-2018, Spring 2010, 2011 Fundamentals of Biology Laboratory (BIOS 120L) Spring 1991- Fall 1998

General Biology (BIOS 101, for non-science majors) Fall 1993-Spring 1998, Spring 2000, 2003, 2007, 2015, Summer 2007, Fall 2008, 2013 General Biology Laboratory (BIOS 101L) Fall 1993-Spring 1994, 2005

General Microbiology, Fall 2000

Human Physiology (BIOS 280), Summer 2007, 2009

Human Heredity and Society (BIOS 106) Spring 1995

Integrated Sciences I (SC 101), team-taught with Cynthia Sisson, Fall 2002, team-taught with Laura Whitlock, Fall 2003 Integrated Sciences II (SCI 102), team-taught with Cynthia Sisson, Spring 1999 Integrated Sciences III (SC 103), team-taught with Wayne Gustavson, Fall 2001, 2002

Seminar (BIOS 495) Fall 1994, 1996, 1998, Summer 2002

Special Topics (BIOS 490) Proteomics and Genomics, Spring 2007, Biology for the 21st Century, Spring 2011, Neurobiology, Fall 2015, 2017

Courses taught as 100% online courses: FS 104, BIOS 120, BIOS 120L, BIOS 210, BIOS 363, BIOS 363L, BIOS 434, BIOS 480

Teaching Experience: Graduate Associate Member of the LSU-S Graduate Faculty

Supervised Paula Polk, MS earned in December 2000, LSU-Shreveport/LSUHSC-S Cooperative Masters Program

600 Level (Since 2011): Advanced Developmental Biology Spring 2015, 2016, 2018; Advanced Immunology Summer 2013, Spring 2015, 2016, 2017, 2019; Advanced Immunology Laboratory, Summer 2013; Advanced Endocrinology, Fall 2015, 2017; Advanced Neurobiology, Fall 2015, 2017; Advanced Parasitology, Fall 2017, 2019; Histology and Histology Laboratory, Summer 2018, 2019

Undergraduate Research Students Supervised (since 2011):

Lydia Angel Scott, Fall 2010 and Spring 2011 Matthew Templeton, Fall 2012 Debra Cordell, Spring 2013 Catie Colvin, Spring 2014 Martin Brigham, Spring 2015

Professional Experience

Co-Instructor, Online Scientific Teaching Institute for LA-SiGMA (Louisiana Alliance for Simulation-Guided materials Applications, (<u>http://lasigma.loni.org</u>). Taught a course with Cynthia Sisson for postdocs and graduated students in scientific teaching, July 2012, June 2013, June 2015.

Regional Field Station Leader for <u>FIRST IV:Faculty Institutes for Reforming Science Teaching</u> <u>for Postdoctoral Research Scholars</u>. First IV is supported by NSF DUE 0817224 to Dr. Diane Ebert-May, Michigan State University), 1/2009 – 10/2013. Facilitated four teaching workshops at the University of Georgia Marine Institute and mentored three teams of postdocs in teaching. Scored videos of teaching using the Reformed Teaching Observation Protocol for the grant 2012-2013.

Professional Development

Gordon Research Conference on Undergraduate Biology Education Research, June 23-28, 2019

2012 POGIL (Process Orientated Guided Learning Inquiry) South Central Regional Workshop, 7/10/12 – 7/12/12, University of Texas at Dallas

NSF Chautauqua Short Course for College Teachers in Biomedical Ethics, taught by Denise M. Dudzinski, PhD, MTS, Assistant Professor, Medical Ethics, Department of Medical History & Ethics, University of Washington School of Medicine, University of Washington, July 12-14, 2004.

FIRST II: FIRST II is a National Dissemination Project supported by NSF DUE 0088847 to Dr. Jan Hodder, Oregon Institute of Marine Biology, and Dr. Diane Ebert-May, Michigan State University, to improve the teaching of science (http://www.first2.org). I organized LSUS Institutional Team, prepared the application forms, and serve as a member of the team under the Louisiana Universities Marine Consortium Field Station Team.

Bioinformatics, VectorNet Workshop for Biology Teaching Faculty, taught by Uwe Hilgert, Biology Media Educator, and Michael O'Brien, High School Educator, Dolan DNA Learning Center, Cold Spring Harbor Laboratory, June 16-20, 2003. I have used exercises and materials from the course in two courses that I have taught this summer and fall, and have instructed two other faculty members in the use of the programs and materials available from the Dolan Learning Center.

IEEE Computer Society Bioinformatics Conference Tutorials: "Perl and Bioperl: Tools for Automated Analysis of Biological Sequence Data" and "Comparative Genomics for Biological Discovery", August 14, 2002

Project Kaleidoscope Summer Institute, "Quantitative Literacy: Everybody's Orphan" and "Assessment in the Service of Student Learning", May 29-June 2, 2002

LaCEPT-Sponsored State-Wide Biology Workshop: "Innovation in Large Lectures - Teaching for Active Learning", Dr. Diane Ebert-May, Louisiana Tech University, Ruston, LA, Jan. 15-16, 1998

Developmental Biology Teaching Laboratory Workshop, Darling Marine Center, University of Maine, July 1-4, 1998.

LaCEPT-Sponsored State-Wide Biology Workshop: "Designing and Running Investigative Laboratories", Louisiana Tech University, Ruston, LA, May 25-28, 1998

LSU Board of Regents/LaCEPT Annual Conferences: Shaping the Future of Undergraduate Science, Mathematics, Engineering and Technology Education, Jan. 1994, 1995, 1996, 1997, Baton Rouge, LA.

Member of the Project Kaleidoscope Faculty for the 21st Century, Class of 1994. I attended the Project Kaleidoscope Invitational Symposium: Faculty for the 21st Century "Creating Our Futures", October 28-30, 1994 in Atlanta, GA and National Assembly of the Project Kaleidoscope Faculty for the 21st Century, Oct. 11-13, 1996, Kansas City, MO.

Invited Participant: "Biology in Action: New Approaches to Teaching and Learning Science", May 22-25, 1996, Radford University, Radford, VA 24142.

Selected by the LSU Board of Regents as a Summer 1996 LaCEPT Faculty Intern. I participated as a Faculty Intern in the LaSIP Earth Science Teacher Enhancement Program at Centenary College, Shreveport, LA.

Grant Support

Member of the General Biology Lecture and Lab Cohort for the Louisiana Library Network Grant for the Interactive Open Educational Resources (OER) . Summer 2021-2022

Louisiana Board of Regents Support Fund, PI: "Strengthening of Student Preparation in the Biological Sciences Through Training in Inquiry and Computational Biology", 2009-2011, \$36,400. Funds for the purchase of machine-vision workstations and the preparation of laboratory exercises that use the workstations and data generated with the workstations. Louisiana Board of Regents Support Fund, PI: "Enhancement of Proteomics and Genomics in the Biological Sciences Curriculum", 2006-2007, \$28,000. Funds for the development of new courses.

North Louisiana Partnerships in Innovation, "Pilot Study to Determine the Feasibility of a New Biotechnology Certificate within the Biology Degree Program", Nov.1, 2004-Oct.31, 2005, \$10,000.

LSU-Shreveport Technology Fee Fund, Equipment and Software for Lecture Capture. 2004-05, \$14,945.

Louisiana Board of Regents Support Fund, PI: "Equipment to Enhance Teaching Genomics in the Biological Sciences Curriculum", 2003-2004, \$64,000. Funds for the upgrading of the Biological Sciences Computer Laboratory.

National Science Foundation MCB 9604180: RUI, PI: PTL-1, A Tau-Like Microtubule-Binding Protein in *C. elegans*, 7/1/97 to 6/30/2002, \$105,000.

Louisiana Board of Regents Support Fund, PI: "Advanced Microscopy Workstations for the Enhancement of Undergraduate Education in Biological Sciences and Integrated Sciences", 2000-01, \$35,000.

Louisiana Board of Regents Support Fund, PI: "Stereomicroscopes for the Enhancement of Upper division Courses in the Biological Sciences", 2000-02, \$70,000.

NSF BIR 9602940, Biological Equipment and Instrumentation Resources Program, PI, A High Speed Centrifuge for Preparation of Biological Materials, 1997-98, \$21,393.

Louisiana Board of Regents Support Fund, Co-PI: Enhancement of Biology and Biochemistry Instruction at LSUS, 1997-1998, \$43,000.

Louisiana Board of Regents Support Fund, Co-PI: Equipment for the Enhancement of Biochemistry and Molecular Biology at LSU-S, 1997-98, \$66,845.

Louisiana Board of Regents Support Fund , Co-PI: Equipment to Facilitate Quantitative Investigations in Introductory Biology Laboratories, 1997-98, \$20,000.

American Heart Association, Louisiana Affiliate, with undergraduate Timothy Chia: Determination of the Loss of Function Phenotype of *C. elegans* ptl-1. \$1600, Summer 1997.

Biomedical Research Foundation of Northwest Louisiana, Co-PI: Equipment to Enhance the Molecular Biology and Biochemistry Teaching and Research at Louisiana State

University-Shreveport, 1996-97, \$16,916.

Service

LSU Shreveport SACS-COC QEP Committee, 2013-2020, QEP Director, 2015-2020 LSU Shreveport Health Sciences Advisory Committee, 1992-LSUS Faculty Senate Instruction and Professional Development Committee, Chair, 2019-LSUS Faculty Senate Policy and Personnel Committee, 2017-2019 LSU Shreveport Financial Aid Appeals Committee, 2013-2018 LSU Shreveport Advising Committee, 2009-2014

Stephen W Banks, M.Sc, Ph.D, C.Biol, FIBiol, MRSC



Title: Professor, Chair of Biology Start Year at LSUS: 1992 Department: Biological Sciences Department Office Location: Science Building 216 Office Phone: 318-797-5220 Office Fax: 318-797-5222 Email: stephen.banks@lsus.edu

Selected Publications:

**Denotes Undergraduate Student, *Denotes Graduate Student

Banks, S.W., Steele, M.J., Ward, D., and Dewick, P.M., (1982) "Stereochemistry of Isoflavone Reduction during Biosynthesis of (+)-and (-)-Pterocarpans: ²H Nuclear Magnetic Resonance Studies of the Biosynthesis of (+)-Pisatin and (-)-Medicarpin". *Journal of the Chemical Society: Chemical Communications* 1982 p.147-149.

Banks, S.W., and Dewick P.M., (1982) "Biosynthesis of the 6a-Hydroxy-Pterocarpans: Deuterium NMR evidence for direct Hydroxylation of Pterocarpans" *Zeitschift Naturforshung* 38c:185-188.

Banks, S.W., and Dewick P.M., (1982) "(-)-Pisatin, an induced metabolite of abnormal configuration from *Pisum sativum*" *Phytochemistry* **21**:1605-1608.

Banks, S.W., and Dewick P.M., (1982) "Biosynthesis of the 6a-Hydroxypterocarpan Phytoalexin Pisatin in *Pisum sativum*" *Phytochemistry* **21**: 2235-2242.

Banks, S.W., and Dewick P.M., (1983) "Biosynthesis of Pisatin: Experiments with Enantiomeric Precursors" *Phytochemistry* **22**:1591-1595.

Banks, S.W., and Dewick P.M., (1983) "Biosynthesis of Glyceollins I, II and III in Soybean (*Glycine max*)" *Phytochemistry* **22**:2729-2733.

Smith, D.A., Wheeler, H.E., Banks, S.W. and Cleveland T.E., (1984) "Association between Lowered Kievitone Hydratase Activity and Reduced Virulence to Bean in Variants of *Fusarium solani* f.sp. *phaseoli*" *Physiological Plant Pathology* **25:**135-147.

Smith, D.A., Banks, S.W., (1986) "Biosynthesis, Elicitation and Biological Properties of Isoflavanoid Phytoalexins". *Phytochemistry* **25**:979-995.

Graves, A.E., Goldman, S.L., Banks, S.W., (1988) "Scanning Electron Microscope Studies of *Agrobacterium tumefaciens*: Attachment to *Zea mays*, *Gladiolus* sp., and *Triticum aestivum*". Journal of Bacteriology **170**:2395-2400.

Banks, S.W., Gossett, D. R., Lucas, M.C., Millhollon, E. P., **LaCelle, M.G., (1993) "Agrobacterium-Mediated Transformation of Kenaf (*Hibiscus cannabinus* L.) with the *b*-Glucuronidase Gene" *Plant Molecular Biology Reporter* **11**:(2)101-104.

Gossett, D.R., Millhollon, E.P., Lucas, M.C., Banks, S.W., **Marney, M-M., (1994) "The effects of NaCl on antioxidant enzyme activities in callus tissue of salt-tolerant and salt-sensitive cotton (*Gossypium hirsutum* L.) cultivars". *Plant Cell Reports* **13**:498-503.

Gossett, D.R., Banks, S.W., Millhollon, E.P., Lucas, M.C.,(1996) "Antioxidant response to NaCl stress in a control and an NaCl-tolerant cotton cell line grown in the presence of paraquat, butathione, sulfoximine and exogenous glutathione" *Plant Physiology* 112:803-809.

*Rainwater, D.T., Gossett, D.R., Millhollon, E.P., Hanna, H.Y., Banks, S.W., Lucas, M.C., (1996) "The relationship between yield and the antioxidant defense system in tomatoes grown under heat stress" *Free Radical Research* **25:**421-435.

Manchandia, A.M., Banks, S.W., Gossett, D.R., *Bellaire, B.A., Lucas, M.C., Millhollon, E.P., (1999) "The Influence of *a*-Amanitin on the Induction of Antioxidant Enzymes during Salt Stress" *Free Radical Research* **30:429-438.

*Rajguru, S.N., Banks, S.W., Gossett, D.R., Lucas, M.C., Fowler, T.E. Millhollon, E.P., (1999)
 Antioxidant Response to Salt Stress During Fiber Development in Cotton Ovules. *The Journal* of Cotton Science 3: 11-21. Online at the Cotton
 Foundation: http://www.jcotsci.org/1999/issue01/phys/art01/article.pdf

*Bellaire, B.A., *Carmody, J., *Braud, J., Gossett, D.R., Banks, S.W., Lucas, M.C., *Fowler, T.E. (2000) Involvement of Abscisic Acid-Dependent and -Independent-Pathways in the Up-regulation of Antioxidant Enzyme Activity During NaCl Stress in Cotton Callus Tissue. *Free Radical Research* **33**: 531-545.

Banks, S.W., Shepherd, D.K., Williams-Hart, T., Gossett, D.R., and Crnkovic, A.C. (2007) "Snail shells in a practical application of statistical procedures". *Journal of Biological Education*. **41:** (3) 131-137.

*Vital, S.A., *Fowler, R.W., *Virgen, A., Gossett, D.R., Banks, S.W., Rodriguez, J. (2008) Opposing Roles for Superoxide and Nitric Oxide in the NaCl-Induced Up-Regulation of Antioxidant Enzyme Activity In Cotton Callus Tissue. *Journal of Experimental and Environmental Botany* **62**: 60-68.

Jones, L., Gossett, D.R., Banks, S.W., McCallum, M.L., (2010) Antioxidant Defense System in Tadpoles of the American Bullfrog *(Lithobates catesbeianus)* Exposed to Paraquat *Journal of Herpetology*, **44: (2) 222–228.

Beverly Burden, PhD



Name: Dr. Beverly J Burden

Title: Associate Professor

Start Year at LSUS: 1994

Department: Biological Sciences Department

Office Location: Science Building 115D

Office Phone: 318-797-5088

Email: <u>beverly.burden@lsus.edu</u>

Matyas Buzgo, PhD



Name: Dr. Matyas Buzgo Title: Associate Professor Start Year at LSUS: 2007 Department: Biological Sciences Department School: Mathematics and Sciences College: Arts and Sciences Office Location: Science Building 115A Office Phone: 318-797-5120 Office Fax: 318-797-5222 Email: matyas.buzgo@lsus.edu

Research Interests:

General and Past:

My research interest is evolutionary biology of land plants, especially flowering plants (angiosperms) and non-flowering seed plants (gymnosperms). I am particularly interested in the evolution of development (EvoDevo) of the flower and floral organs, and other reproductive organs. Where did carpels and stamens come from? How did the ancestral organs look and how were they arranged throughout the plant, before we would recognize them as "flower"? Do we have fossil representations of these pre-angiosperms? Apart from the sporangia, what is the principal difference between a vegetative leaf and a floral organ? Is there any, at all?

Actual, and Student Research Opportunities:

Synopsis:

1) Leaf development and evolution in Eusporangiate ferns: Ophioglossaceae and Psilotaceae are odd siblings. We re-examined the morphology of Psilotum. and found that the "green aerial frond" corresponds rather to fern aleaf than to a stem. With this, Psilotum indeed is "similar" to Ophioglossaceae in terms of organ identity of its parts. The differences that meks these taxa so oddly dissimilar are based on differences of their development: Heterochony and Homeotisis.

2) Fern embryology: tip-to-toe embryo

In the very early stage, also the embryo of regular ferns (model organism Ceratopteris) develops in a linear, bipolar manner. However, we have not yet identified hypostasis, root initiation in Ceratopteris, and we still have a long way to study the embryo of Pslotum or Ophioglossaceae.

3) Plant tissue culture: Louisiana Iris, day lily

The purpose is to establish a successful protocol for plant tissue cultures derived from shoot apex meristems or parenchyma cells of select organism, such as Iris and Hemerocallis.

Details:

1) Leaf development and evolution in Eusporangiate ferns

Previous Team: Annie Phanid Miller, Sarah Witherington, Cassie Cole, Dr. Cran M Lucas; BSA 2010 242, 244.

"Ferns" in general (Monilophytes) are the closest relatives of extant seed plants, and many know them as an archaic group of plants, associated with dinosaurs, etc. However, most ferns ("ferns" in the narrow sense) belong to a single group (or "clade") called Leptosporangiate Ferns, based on their very peculiar sporangium. This group radiated relatively recently, and is not really "archaic", but actualy modern; sadly, most truely archaic groups have died out. From all the "old style" ferns (eusporangiate ferns), only three groups have survived to today: Horsetails (Equisetaceae), Boat-ferns (Marrattiaceae), and the group of Ophioglossaceae and Psilotaceae.

This last group, Ophioglossaceae and Psilotaceae, is the biggest surprise of the last years' research. Psilotaceae, seemingly without leaves and or roots, was long time considered not a fern, but actually older than all other vascular land plants, older even than club-mosses (Lycophytes). Ophioglossaeae, however, with a peculiar stem growth, vascular system, and an elaborate axial, stem-like outgrowth bearing the sporanga, has been considered closer related to seed plants than normal ferns. Yet, molecular analysis revealed these two "opposite" families as two monophyletic sisters, and within the monilophytes. And: while a review of morphological data showed a great correspondance to molecular findings, it could not copy the molecular data for the Ophioglossaceae-Psilotaceae clade.

We re-examined the morphology of Psilotum. and found that the "green aerial frond" corresponds rather to fern aleaf than to a stem. With this, Psilotum indeed is "similar" to Ophioglossaceae in terms of organ identity of its parts. The differences that meks these taxa so oddly dissimilar are based on differences of their development: Heterochony and Homeosis.

2) Fern embryology: tip-to-toe embryo in c-fern

Previous Team: Christopher Young, Jonette Green, Lauren Woodward.

Psilotum realy has no root - what happened to it? Is it initiated in the embryo? Actually, how do ferns in general initiate roots?

Seed plants have a strict bi-polarity in the pro-embryo: suspensor-embryo. The rood forms in below the suspensor from a defined cell layer (the "hypostatsis"), and grows clearly into the direction opposite to the shoot apex of the embryo. However, in fern embryos we knew, root and shoot apex emerge in one direction, opposite to the suspensor. That is, even the root development of regular ferns is not really understood.

The leptosporangiate fern Ceratopteris ("c-fern") is a model organism for fern genetics and development. We established a protocol for its gametophyte cultivation and were able to study the very early embryo development. We found, that in this very early stage, also the fern embryo develops in a linear, bipolar manner. However, we have not yet identified hypostasis, root initiation in Ceratopteris, and we still have a long way to study the embryo of Pslotum or Ophioglossaceae.

3) Plant tissue culture: Louisiana Iris, day lily

The project is in collaboration between LSUS (Biological Sciences) and Plantation Point Nursery (Mooringsport LA). The organism are Louisiana iris and day lilies (Iris section Apogon and Hemerocallis). The primary intent is to transfer rhizomatous tissue (parenchyma and shoot apex meristems) to a solid growth gel containing cell dedifferentiating growth factors, resulting in a plant tissue callus. A second approach may use seeds as tissue start.

The bulk of storage and maintenance operation would be in Mooringsport and not burden LSUS facilities. It provides the opportunity to train students on a standard method used in agriculture, horticulture, and plant biology. The outlook of this project is to apply the findings onto the fern research project mentioned above.

Selected Publications:

Haiying, L., Carlson, J.E., Leebens-Mack, J.H., Wall,P.K., Mueller, L.A., Buzgo, M., Landherr, L.L., Hu, Y., DiLoreto, D.S., Ilut, D.C., Field, D., Tanksley, S.D., Ma, H. dePamphilis, C.W. 2007. An EST Database for Liriodendron tulipiferaL. floral buds: the first EST resource for functional and comparative genomics in Liriodendron. Tree Genetics and Genomes 4(3): 419-433.

Soltis, D.E., Chanderbali, A.S., Kim, S., Buzgo, M., Soltis, P.S. 2007. The ABC model and its applicability to basal angiosperms. Annals of Botany 100: 155-163.

Buzgo, M., Chanderbali A.S., Kim, S., Zheng, Z., Oppenheimer, D., Soltis, P.S., Soltis, D.E. 2007. Floral developmental morphology of Persea americana (avocado, Lauraceae): the oddities of male organ identity. International Journal of Plant Sciences 168 (3): 261-284.

Buzgo, M., Soltis, D.E., Soltis, P.S., Kim, S., Ma, H., Hauser, B.A., Leebens-Mack, J., Johansen,
B. 2006. Perianth development in the basal monocot Triglochin maritima (Juncaginaceae).
- In: Columbus, J.T., Friar, E.A., Porter, J.M., Prince, L.M., Simpson, M.G. (eds), Monocots:
Comparative Biology and Evolution, (excluding Poales). Claremont, CA, USA: Rancho Santa
Ana Botanic Garden, pp. 107-125. (see also Aliso 22).

Soltis, P.S., Soltis, D.E., Kim, S., Chanderbali, A., Buzgo, M. 2006. Expression of floral regulators I basal angiosperms and the origin and evolution of ABC function. - In: Soltis, D.E., Leebens-Mack, J.H., Soltis, P.S. (eds), Advances in Botanical Research, Vol. 44, Devlopmental Genetics of The Flower. Elsevier Ltd., London, UK (Academic Press, San Diego CA, USA), pp. 483-506.

Albert, V.A., Soltis, D.E., Carlson, J.E., Farmerie, W.G., Wall, P.K., Ilut, D.C., Solow, T.M., Mueller, L.A., Landherr, L.L., Hu, Y., Buzgo, M., Kim, S., Yoo, M.-J., Frohlich, M.W., Perl-Treves, R., Schlarbaum, S., Bliss, B.J., Zhang, X., Tanksley, S., Oppenheimer, D.G., Soltis, P.S., Ma, H., dePamphilis, C.W., Leebens-Mack, J.H. 2005. Floral gene resources from basal angiosperms for comparative genomics research. BMC Plant Biology 5 (1): 5.

Buzgo, M., Soltis, P.S., Kim, S., Soltis, D.E. 2005. The making of a flower. The Biologist 52: 149-154.

Buzgo, M., Soltis, P.S., Soltis, D.E. 2004. Floral developmental morphology of Amborella trichopoda (Amborellaceae). International Journal of Plant Sciences 165: 925-947.

Buzgo, M., Soltis, D.E., Soltis, P.S., Ma, H. 2004. Towards a comprehensive integration of morphological and genetic studies of floral development. Trends in Plant Science 9: 164-173.

Personal Bio:

Associate Professor: Department of Biology, Louisiana State University in Shreveport, Shreveport, LA, USA 2007-present

Postdoctoral experience:

Department of Botany, University of Florida, Gainesville, FL, USA (with Drs. Douglas E. Soltis and Pamela S. Soltis) 2002-2007 Dept. of Evolutionary Botany, University Copenhagen, Denmark (with Dr. Bo Johansen) winter 2001 Jodrell Laboratory, Royal Botanic Gardens, Kew, UK (with Drs. Mark W. Chase and Paula J. Rudall) 2000-2001 Institute of Systematic Botany, University of Zurich 1999 **Other:** System Engineer, IBM Switzerland Informatica, 1992-1994 Instructor and field guide, WWF Switzerland, 1989-1994 Laboratory technician, public relations officer and statistical analyst, Health Department of the City of Zurich, 1989-1992 Teaching Assistant, Research Assistant, University of Zurich, 1989-1992 and 1994-1999

Amy Anne Erickson, PhD



Associate Professor, Biological Sciences, LSUS Graduate Director, MS Biological Sciences Program Department of Biological Sciences Phone (318) 797-5105 amy.erickson@lsus.edu

Education:

1996-2003 Ph. D. in Biology, University of South Florida (Advisors: Susan S. Bell, Clinton J. Dawes), "Associational resistance from and susceptibility to herbivory by the mangrove tree crab, *Aratus pisonii*, on the red mangrove, Rhizophora mangle when in the presence of the black mangrove, *Avicennia germinans*, and the white mangrove, *aguncularia racemosa*"

1994-1997 M. S. in General Experimental Psychology, Florida State University (Advisor: Richard L. Hyson), "Characterization of the GABA_A receptor in the chick brainstem auditory system"

1990-1994 B. S. in Biology, Georgetown University (Advisor: Philip Sze), Senior Thesis: "The effects of herbivory on macroalgae: an amphipod preference feeding study"

Professional Experience:

	1
2019-2021	Graduate Director, Master in Biological Sciences Program, LSUS
2013-	Associate Professor, Dept. of Biological Sciences, Louisiana State University
	Shreveport, (12-15 hr teaching load/semester); Granted tenure
2012-2016	Member, Science & Education Advisory Council, LUMCON
2007-2013	Assistant Professor, Dept. of Biological Sciences, Louisiana State University
	Shreveport, (9-12 hr teaching load/semester)
2006-	Research Associate/Collaborator, Smithsonian Institution
2006-2007	Visiting Assistant Professor, Dept. of Integrative Biology, University of South
	Florida (6 hr teaching load/semester)
2006	Postdoctoral Fellow, Smithsonian Marine Station, FL (Advisor: Valerie J.
Paul) "Iwnah	va blooms in the Indian River Lagoon"

Paul), "Lyngbya blooms in the Indian River Lagoon"

2004-2005	Caribbean Coral Reef Ecosystems Postdoctoral Fellow, Smithsonian Marine Station, FL (Advisors: Valerie J. Paul, Ilka C. Feller), "Are feeding
	preferences of the mangrove tree crab Aratus pisonii
chemically	determined?"
2003-2004	Postdoctoral Fellow, Smithsonian Marine Station, FL (Advisor: Valerie J.
Paul),	"Examination of whether coral reef herbivore feeding preferences are
	chemically based"
2002-2003	Adjunct Instructor, Dept. of Biology, University of Tampa
1996-2003	Instructor & Graduate Teaching Assistant, Dept. of Biology, University of
South	Florida
1994-1996	Graduate Teaching & Departmental Assistant, Dept. of Psychology, Florida
State	University
1994	Undergraduate Teaching Assistant, Dept. of Biology, Georgetown University

Submissions and Publications:

 Capper, A., Erickson., A.A., Ritson-Williams, R., Becerro, M.A., Arthur, K.A., and Paul,
 V.J. (2016). Palatability and chemical defences of benthic cyanobacteria to a suite of herbivores. J. Exp. Mar. Biol. Ecol. 474:101-108.

Erickson, A.A., Bell, S.S., and Dawes, C.J. (2012). Associational resistance protects mangrove leaves from crab herbivory. Acta Oec. 41:46-57.

Erickson, A.A., Feller, I.C., Paul, V.J., Kwiatkowski, L.M., and Lee, W. (2008). Selection of an omnivorous diet by the mangrove tree crab *Aratus pisonii*. J. Sea Res. 59:59-69. Erickson, A.A., Paul, V. J., Van Alstyne, K.L., and Kwiatkowski, L.M. (2006). Palatability of green algae that may employ different types of activated chemical defenses. J. Chem.

Ecol. 32:1883-1895.

Ellis, William L., Bowles, Justin W., Erickson, Amy A., Stafford, Nate, Bell, Susan S., and Thomas, Melanie. (2006). Alteration of the chemical composition of mangrove (*Laguncularia racemosa*) leaf litter fall by freeze damage. Estuar. Coast. Shelf. Sci. 68:363-371.

Erickson, A.A., Bell, S.S., and Dawes, C.J. (2004). Does mangrove leaf chemistry help explain crab herbivory patterns? Biotropica 36(3):333-343.

Erickson, A.A., Saltis, M., Bell, S.S., and Dawes, C.J. (2003). Herbivore feeding preferences as measured by leaf damage and stomatal ingestion: a mangrove crab example. J. Exp. Mar. Biol. Ecol. 289(1):123-138.

In preparation:

Burks, E., Stratton, C. and Erickson, A.A. (in preparation). The effect of freshwater acidification on freshwater clams.

Sanson, W., Burden, B., and Erickson, A.A. (in preparation). Anti-microbial effects of *Salvinia molesta*.

Erickson, A.A., Paul, V.J, Kwiatkowski, L.M., and Becerro, M.A. (in preparation). Sea urchin species vary in their susceptibility to algal chemical defenses.

Erickson, A.A., Bell, S.S., and Dawes, C.J. (in preparation). Influence of mangrove species associations on herbivory by mangrove tree crabs.

Erickson, A.A., Feller, I.C., Paul, V.J., Kwiatkowski, L.M., West, L., and Trejo, P. (in preparation). The impact of leaf age and pre-existing crab damage on feeding preferences by the mangrove tree crab *Aratus pisonii*.

Erickson, A.A., Feller, I.C., Paul, V.J., Kwiatkowski, L.M., Lee, W. (in preparation).

Chemical mediation of feeding preferences for the mangrove tree crab Aratus pisonii.

- Erickson, A.A., Parsons, K., Feller, I.C., and Lee, W. (in preparation). Characterization of *Aratus pisonii* populations in Indian River Lagoon mangrove impoundments post Hurricanes Frances and Jeanne.
- Erickson, A.A., Sargeant, B., Holstein, H., Lincoln, A., and Bell, S.S. (data analysis). Time budget analysis of the mangrove tree crab *Aratus pisonii*.

Grants, Honors, and Awards:

2021	LSUS Tech Fee, Erickson, A. Supplying biology labs with compound light microscopes. Awarded: \$34,500
2021	BoRSF Departmental Enhancement Grant, Erickson, A. (PI), Siska, P., Chen,
2021	X., Joiner, G., Maguigan, M., Providing GIS training to the LSUS community and Northwest Louisiana. Awarded: \$61,400
2021	LSUS Faculty Research and Development Grant, Erickson, A., Supplies for project on environmental DNA. Awarded: \$5,000
2020	LSUS Tech Fee, Erickson, A. Resources for environmental DNA research. Awarded: \$2,870
2020	LSUS Tech Fee, Mast J., (Advisor: Erickson, A.A.), Supplying biology labs with stereomicroscopes. Awarded: \$16,500
2020	LSUS Faculty Research and Development Grant, Erickson, A., Supplies for project on environmental DNA. Awarded: \$5,000
2020	LSUS Faculty Research and Development Grant, Erickson, A., Travel for environmental DNA field studies. Awarded: \$1,500
2020	LSUS Faculty Research and Development Grant, Erickson, A., Supplies for project on acidification of freshwater and marine ecosystems. Awarded: \$1,500
2019	LSUS Tech Fee, Erickson, A., Chen, X., Lewis, J. Resources for geographic information science. Awarded: \$21,990
2019	LSUS Tech Fee, Mast J., (Advisor: Erickson, A.A.), Laptop chargers for use
in	the classroom. Awarded: \$1,890
2019	LSUS Faculty Research and Development Grant, Erickson, A., Funding for research assistant for BoRSF grant. Awarded: \$5,000
2018	LSUS Research Sabbatical: Fall 2018
2018	LSUS Tech Fee, Erickson, A., Preparing plant samples for CHN analysis, Awarded: \$5000
2018	BoRSF Research Competitiveness Subprogram Grant – One-year, Erickson,
A., Allelo	pathic effects of common aquatic plants on the invasive water fern Salvinia molesta. Awarded: \$15,000
2017	LSUS Tech Fee, Garcia S., (Advisor: Erickson, A.A.), Video cameras for use
in	the classroom and research. Awarded: \$5,244

2017	LSUS Faculty Research and Development Grant, Erickson, A., Generating
plant	extracts for chemical ecology experiments for use in research and in
the	classroom. Awarded: \$1000.
2017	Campus Federal's Teaching Enhancement Fund, Erickson, A., Attending
	Southern Section-American Society of Plant Biologists Meeting.
Award	ed: \$200
2016	LSUS Faculty Research and Development Grant, Erickson, A., Presentation
of	Scientific Research at the 4 th Mangrove Macrobenthos Conference.
Award	-
2016	BoRSF Undergraduate Enhancement Grant, Erickson, A. (PI), Burden, B.
	Buzgo, M., Gossett, D., Enhancement of Environmental Science Equipment at
	LSUS. Awarded: \$62,800
2015	Campus Federal's Teaching Enhancement Fund, Erickson, A., Attending
	Teaching Online Conference, Awarded: \$250
2014-	Harman and Renae Chandler Endowed Professorship for the Master of
	Biological Science Program
2014	LSUS Tech Fee, Erickson, A., Supplying Biology Laboratories with a
	Stereomicroscope with an Integrated Camera System, Awarded: \$2,600
2014	LSUS Tech Fee, Erickson, A., Supplying Biology Laboratories with Van
	Dorn Alpha Jars for Water Sampling, Awarded: \$1042.28
2013	Campus Federal's Teaching Enhancement Fund, Erickson, A., Attending
	Science Online: Oceans Conference, Awarded: \$300
2013	LSUS Tech Fee, Erickson, A., Supplying Biology Laboratories with
	a Stereomicroscope with an Integrated Camera System, Awarded: \$2500
2013	LSUS Tech Fee, Erickson, A., Supplying Biology Laboratories with Van
	Dorn Alpha Jars for Water Sampling, Awarded: \$984
2012	LSUS 2012 Tech Fee, Erickson, A., Supplying Biology Laboratories with
	Stereomicroscopes & Integrated Camera Systems, Awarded: \$14,727
2012	BoRSF Undergraduate Enhancement Grant, Erickson, A., Banks, S. Gossett,
D.,	Lucas, C., Enhancement of Analytical Skills in Chemical Ecology.
	Awarded: \$37,824
2011	LSUS Tech Fee, Erickson, A.A., Supplying Biology laboratories with
	microscopes. Awarded \$22,500
2010	NSF-MRI, Lucas, C., Gossett, D., Banks, S., Williams-Hart, T., Erickson, A.
	(Co-PI), Acquisition of a High Performance Liquid Chromatography (HPLC) System
	for Research, Research Training and Education. Awarded: \$152,692
2010	LSUS Tech Fee, Morlock, T., and Key. M., (Advisor: Erickson, A.A.),
	Increasing drying capacity of the lyophilizer. Awarded: \$2,202
2010	LSUS Tech Fee, Erickson, A.A., and Banks, S.W., Supplying Biology
• • • • •	laboratories with balances. Awarded: \$4,810
2010	BoRSF Traditional Enhancement Grant, Erickson, A.A., To the sea and back:
	Building a bridge between Louisiana State University Shreveport (LSUS) and
2000	Louisiana Universities Marine Consortium (LUMCON). Awarded: \$5,465
2009	LA EPSCoR Pfund Grant, Erickson, A.A., Effect of nutrient fertilization on
	nutritional and defensive chemistry of mangrove leaves in Florida and Belize.
	Awarded: \$10,000 (plus \$2,378 as in-cash match)

2008	LSUS Faculty Development Grant, Erickson, A.A., Request for travel to the
	Benthic Ecology Meeting. Awarded: \$500
2008	LSUS Faculty Research Grant, How mangrove leaf chemistry influences
feeding	g by the mangrove tree crab Aratus pisonii. Awarded: \$597
2007	University of South Florida Faculty Research and Development Grant, The
role of	chemistry in feeding by marine herbivores. Awarded: \$800
2006-2007	Caribbean Coral Reef Ecosystems Grant, Erickson, A.A., Paul, V.J., and
Feller,	.C., Effect of fertilization on nutritional and defensive chemistry of mangrove
	leaves. Awarded: \$1000
2003	Student Travel Award, Estuarine Research Federation
2001	Tharpe Summer Fellowship in Biology, University of South Florida
2000	Best Student Presenter, Southeastern Estuarine Research Society
2000	Phi Kappa Phi Chapter Fellowship (126), University of South Florida
2000-2001	Tharpe Fellowship in Biology, University of South Florida
1998-2001	Biology Travel Fund, University of South Florida
1997	Nomination-Provost's Award in Outstanding Teaching, University of South
	Florida
1994	Dean's List, Georgetown University

Teaching Experience:

2013-	Associate Professor, Dept. of Biological Sciences, Louisiana State University Shreveport: same courses as below; Graduate Biostatistics (F2F & online), Erechmen Seminar Investive Species, Ten down & Bettern un Effects, Marine
	Freshman Seminar, Invasive Species, Top-down & Bottom-up Effects, Marine Feeding Ecology Lab, <i>Salvinia</i> , Field and Organismal Discussion Group; Total preps: 20
2007-2013	Assistant Professor, Dept. of Biological Sciences, Louisiana State University Shreveport: General Biology (non-science majors; F2F & online), Marine Environment (non-science majors; online), Biological Inquiry (F2F & online), Principles of Ecology & Lab (F2F & online), Aquatic Biology (F2F & online) & Lab, Marine Biology (F2F & online) & Lab, Chemical Ecology (F2F & online) & Lab, Marine Feeding Ecology, Applied Coastal Ecology (online); Team teach for 1 wk/sem: Biology Principles I Lab, Research Methods, Medical Seminar. Total preps: 13
2006-2007	Visiting Assistant Professor, Dept. of Integrative Biology, University of South Florida: Principles of Biology (non-science majors), Marine Biology (non-science majors; online), Marine Biology (for majors), Advanced Marine Biology, 2006 Invited Lecture, Semester at Sea, Florida Atlantic University/HBOI: Algal chemical defenses. Total preps: 4
2003	Mangrove Ecology Scientist, Electronic field trip, Smithsonian Environmental Research Center: Where the Land and Sea Intertwine
2002-2003	Adjunct Instructor, Dept. of Biology, University of Tampa: Environmental Science
2002-2003	Lecture Assistant, Dept. of Biology, University of South Florida: Principles of Ecology
2000-2002	Instructor, Dept. of Biology, University of South Florida: Environment

Science Instructor, Fledge-ling Camp of Geology and Ecology (NSF),
of South Florida
Teaching Assistant, Dept. of Biology, University of South Florida:
Laboratories for Principles of Ecology (Head TA), Introductory
& 2
Teaching Assistant, Dept. of Psychology, Florida State University:
Sensation & Perception Laboratory
Teaching Assistant, Program in Medical Sciences, Florida State University:
Neuroanatomy of the Human Brain Laboratory
Teaching Assistant, Dept. of Biology, Georgetown University: Introductory
Biology 2 Laboratory

Contributed Papers:

2015 Southeastern Estuarine Research Society Meeting, Jacksonville, FL: Erickson, A.A., Bell, S.S., and Dawes, C.J., Associational resistance protects mangrove leaves from crab herbivory. 2006 Ecological Society of America, Memphis, TN: Erickson, A.A., Feller, I.C., Paul, V.J., and Kwiatkowski, L.M., Chemical mediation of feeding preferences for the mangrove tree crab Aratus pisonii. 2006 Meeting of the Mangrove Macrobenthos II, Coolangata, Australia: Erickson, A.A., Feller, I.C., Paul, V.J., and Kwiatkowski, L.M., Chemical mediation of preferences for the mangrove tree crab Aratus pisonii. feeding 2006 Southeastern Estuarine Research Society Meeting, St. Augustine, FL: Erickson, A.A., Paul, V.J., Van Alstyne, K.L., and Kwiatkowski, L.M., Palatability of green algae that employ different types of activated chemical defenses. 2004 Ecological Society of America, Portland, OR: Erickson, Amy A., Bell, Susan S., and Dawes, Clinton, J., Relating mangrove herbivore preference and consumption to leaf chemistry and structure. 2004 Association of Tropical Biology and Conservation, Miami, FL: Erickson, Amy A., Bell, Susan S., and Dawes, Clinton, J., Relating mangrove herbivore and consumption to leaf chemistry and structure. preference Benthic Ecology Meeting, Orlando, FL: Erickson, Amy A., Saltis, M., Bell, Susan 2002 S., and Dawes, Clinton J., Dietary preference for red mangrove leaves as measured by leaf damage and crab gut contents. Florida Ecological and Evolutionary Symposium, Archbold, FL: Brooks, R.A., 2000 Bell, S.S., Ellis, W.L., Erickson, A.A., Jacobson, S.B., and Rizzuto, N.V., Ecological engineers: revisiting the past and evaluating the future. Southeastern Estuarine Research Society, Tampa, FL: Erickson, Amy A., 2000 Bell, Susan S., and Dawes, Clinton J., Crab herbivory on the red mangrove Rhizophora mangle L. when found in pure stands versus when in association with the black mangrove Avicennia germinans (L.) Stearn and the white mangrove Laguncularia racemosa (L.) Gaertn. 2000 Benthic Ecology Meeting, Wilmington, NC: Erickson, Amy A., Bell, Susan S., and Dawes, Clinton J., Crab herbivory on the red mangrove Rhizophora mangle L. when found in pure stands versus when in association with the

	black mangrove Avicennia germinans (L.) Stearn and the white mangrove
	Laguncularia racemosa (L.) Gaertn.
2000	Benthic Ecology Meeting, Wilmington, NC: Brooks, R.A., Bell, S.S., Ellis, W.L.,
	Erickson, A.A., Jacobson, S.B., and Rizzuto, N.V., Ecological engineers:
	revisiting the past and evaluating the future.
	To the first man of memory of the second of
Contri	ibuted Posters (*presenter if not first author):
	Entomological Society of America, St. Louis, MO: Burden, B., Sanson Dobbins
Fields,	
1 10103,	W., Erickson, A., Antimicrobial properties of Salvinia molesta on common
	bacteria.
2019	American Society of Plant Biologists, San Jose, CA: Erickson, A, Zittrauer, C., Mast,
2019	
	J., Mulling S. Deen F. Miller P. Owang K. De Sete A. Controlling Scheinig
	Mullins, S., Dean, E., Miller, R., Owens, K., De Soto, A. Controlling Salvinia
2010	<i>molesta</i> through allelopathy.
2019	American Society of Plant Biologists, San Jose, CA: Burden, B., Sanson Dobbins
Fields,	
	W., Erickson, A.*, Antimicrobial properties of Salvinia molesta on common
	bacteria.
2019	American Society of Plant Biologists, San Jose, CA: Mast, J., Mullins, S., Zittrauer,
С.,	
	Dean, E., Erickson, A., Controlling Salvinia molesta through allelopathy.
2019	American Society of Plant Biologists, San Jose, CA: Mast, J., Erickson, A., Burden,
В.,	
	Invertebrate diversity in Salvinia molesta.
2019	4 th Annual LSUS Student Scholars Forum, Shreveport, LA: Mast, J., Mullins, S.,
	Zittrauer, C., Dean, E., Erickson, A., Controlling Salvinia molesta through
	allelopathy.
2019	4 th Annual LSUS Student Scholars Forum, Shreveport, LA: Mast, J., Erickson, A.,
	Burden, B., Invertebrate diversity in Salvinia molesta.
2019	4 th Annual LSUS Student Scholars Forum, Shreveport, LA: Zittrauer, C., Mast, J.,
	Mullins, S., Dean, E., Miller, R., Owens, K., De Soto, A., Erickson, A.,
	Controlling <i>Salvinia molesta</i> through allelopathy.
2019	4 th Annual LSUS Student Scholars Forum, Shreveport, LA: Garcia, S., Bates, A.,
Black,	r Annual 1969 Student Scholars Forum, Snieveport, 174. Sureta, 5., Butes, 14.,
Diack,	B., Graham, L., Greer, K., Mast, J., Erickson, A., Stingray feeding preferences
	in captivity.
2019	4 th Annual LSUS Student Scholars Forum, Shreveport, LA: Garcia, S., Bates, B.,
	4 Annual LSOS Student Scholars Forum, Sineveport, LA. Garcia, S., Bates, B.,
Black,	D. Carlow, J. Carrow, K. Mart, J. England, A. Karatana, Davidstan, in the
	B., Graham, L., Greer, K., Mast, J., Erickson, A., Keystone Predators in the
2010	Rocky Intertidal.
2018	47 th Annual Benthic Ecology Meeting, Corpus Christi, TX: Mast, J., Greer, S.,
Ericks	
	s in the Gulf of Mexico.
2017	Southern Section-American Society of Plant Biologists, Orlando, FL: Whorton, S.,
Mast,	J., Kong, K., Austill, B., Khan, B., Erickson*, A., Possible suppression

of Salvinia molesta by allelopathy of two waterlilies, Nymphaea mexicana and Nymphaea odorata. 2017 Southern Section-American Society of Plant Biologists, Orlando, FL: Mast, J., Whorton, S., Garcia, S., Demarest, A., Erickson, A., Potential allelopathic effects of *Eichhornia crassipes* and *Lemna gibba* on the invasive water fern Salvinia molesta. Southern Section-American Society of Plant Biologists, Orlando, FL: Garcia, S., 2017 Erickson, A., Littoraria irrorata preference for salt marsh habitat. 2nd Annual LSUS Student Scholars Forum, Shreveport, LA: Garcia, S., Erickson, A., 2017 Littoraria irrorata preference for salt marsh habitat. Mangrove Macrobenthos Meeting IV, St. Augustine, FL: Erickson, A.A., Paul, V.J., 2016 Feller, I.C., Lee, W., Kwiatkowski, L.M., West, L., Trejo, P., influence feeding activity by mangrove tree crabs in Mangrove extracts Florida and Belize. 2016 1st Annual LSUS Student Scholars Forum, Shreveport, LA: Al-wadi, A., Zedian, K., Miller, D., Bardwell, C., Candler, M., Mast*, J., Erickson, A., How distance to water affects insect diversity. 2016 1st Annual LSUS Student Scholars Forum, Shreveport, LA: Hood, B., Langston, D., Fuller, K., Patterson*, C., Keyvan, G., Ali, N., Brown, R., Crooks, S., Mast, J., Erickson, A., Insect diversity measured across habitat type. 2016 1st Annual LSUS Student Scholars Forum, Shreveport, LA: Anderson, A., Alvarez, Gomez, T., Jordan, C., Wren, K., Rao, K., Streets, C., Williamson, K., М., Mast*, J., Erickson, A., Influence of Flooding on Insect Diversity in Northwest Louisiana. 2016 1st Annual LSUS Student Scholars Forum, Shreveport, LA: Whorton*, S., Stinson, D., McKay, L., Simmons, A., Ali, H., Mast, J., Erickson, A., The relationship between insect diversity, temperature, and humidity. 2015 Coastal & Estuarine Research Federation, Portland, OR: Erickson, A., Lee, W. habitat use by the mangrove tree crab Aratus pisonii. Trends in 2015 LSUS School of Mathematics and Science Student Research Forum, Shreveport, LA: Hogrefe, K., Wells, D., Russell, K., Spataro, K., Lohman, M., Abubakar*, T., Mast, J., Erickson, A., The relationship Farooqui*, F., between fire ant nest density and environmental factors. 2015 LSUS School of Mathematics and Science Student Research Forum, Shreveport, LA: Cockerham, E., Vagrin, M., Schopp, J., Abubakar, T., Farooqui, F., Mast, J., Erickson, A., Spatial dispersion of fire ant (Solenopsis Vargas, B., invicta) nests. 2015 LSUS School of Mathematics and Science Student Research Forum, Shreveport, LA: Turner, T., Adegboyega, A., Yousuf*, Q., Williams, T., Mast, J., Insect diversity in aquatic habitats. Erickson, A., LSUS School of Mathematics and Science Student Research Forum, Shreveport, LA: 2015 McFarland, D., Cannon, S., Farley, M., Gladney, S., Whorton, S., Erickson, A., How water depth & abiotic factors relate to Mast, J., plankton density & diversity. 2015 Southern Section-American Society of Plant Biologists, Dauphin Island, AL: Erickson. A.A., Bell, S.S., and Dawes, C.J., presenters: Harris, S.*,

Greene, S.*, Associational resistance protects mangrove leaves from crab herbivory. 2015 Southern Section-American Society of Plant Biologists, Dauphin Island, AL: Harris, S., Greene, S., Murcia, H., Mast, J., Whidden, L., Cartee, G., Folkerts, M., A.A., The Determination of Herbivory on Croton capitatus at Erickson. Dickson Park Shreveport, Louisiana. C. Bickham 2015 Southern Section-American Society of Plant Biologists, Dauphin Island, AL: Folkerts, M., Greene, S.*, Harris, S., Mast, J., Fox, J., Jones, A., Erickson, A.A., Characterization of salt marsh habitat surrounding LUMCON's DeFelice Marine Center in Cocodrie, Louisiana. 2015 Southeastern Estuarine Research Society Meeting, Jacksonville, FL: Edwards, M.R., Lincoln, A., Holstein, H., Erickson, A.A., Behavior of the mangrove during its active season. tree crab 2014 LSUS School of Mathematics and Science Student Research Forum, Shreveport, LA: Harris, S., Greene, S., Murcia, H., Mast, J., Whidden, L., Cartee, G., Erickson, A.A., The Determination of Herbivory on Croton capitatus Folkerts, M., at C. Bickham Dickson Park Shreveport, Louisiana. 2014 LSUS School of Mathematics and Science Student Research Forum, Shreveport, LA: Foster, D., Juell, E., Merry, A., Nguyen, B., Wells, C., Wheeler, M., Mast, J.*, Erickson, A.A., The Impact of Human Population Folkerts, M., Density on Bird Diversity in Shreveport, Louisiana. 2014 LSUS School of Mathematics and Science Student Research Forum, Shreveport, LA: Hibbs, B., Jones, K., Ashton, S., Falade, I., Shihab, I., Folkerts, M.*, Erickson, A.A., How does bird diversity differ under variation in land use? LSUS School of Mathematics and Science Student Research Forum, Shreveport, LA: 2014 Shams, S., Harris, H., Simmons, G., Nyajro, E., Folkerts, M., Erickson, A.A., The relationship between native and invasive bird species in Shreveport, Louisiana. 2014 LSUS School of Mathematics and Science Student Research Forum, Shreveport, LA: Barmes, A., Hughes, M., Tanga, S., Ferrell, D., Moore, R., Hubbard, S.*, Folkerts, M., Erickson, A.A., The influence of vegetation G., Shams, and aquatic habitat on bird diversity. 2014 LSUS School of Mathematics and Science Student Research Forum, Shreveport, LA: Folkerts, M., Harris, S., Mast, J., Fox, J., Jones, A., Erickson, A.A., Characterization of salt marsh habitat surrounding LUMCON's **DeFelice** Marine Center in Cocodrie, Louisiana. 2013 BOTANY 2013, New Orleans, LA: Erickson, A.A., Feller, I.C., Paul, V.J., L.M., Lee, W., Selection of an omnivorous diet by the mangrove tree Kwiatkowski. crab Aratus pisonii in laboratory experiments. 2013 BOTANY 2013, New Orleans, LA: Erickson, A.A., Paul, V.J., Kwiatkowski, L.M., Becerro, M., Rhizophytic green algal extracts differentially affect feeding by sea urchin species.

2013 LSUS School of Mathematics and Science Student Research Forum, Shreveport, LA: Folkerts, M., Mast, J., Greene, S., Lincoln, A., Erickson, A.A., Do perform diel vertical migrations in Old River Lake? zooplankton 2013 LSUS School of Mathematics and Science Student Research Forum, Shreveport, LA: Greene, S., Folkerts, M., Mast, J., Lincoln, A., Erickson, A.A., Lakes Do they differ in zooplankton density and diversity? vs. Rivers: 2013 LSUS School of Mathematics and Science Student Research Forum, Shreveport, LA: Mast, J., Greene, S., Folkerts, M., Lincoln, A., Erickson, A.A., The between abiotic factors and zooplankton density in aquatic relationship ecosystems. 2013 Southeastern Estuarine Research Society (SEERS), Charleston, SC: Erickson, A.A., Feller, I.C., Paul, V.J., Kwiatkowski, L.M., Lee, W., Selection of an diet by the mangrove tree crab Aratus pisonii in laboratory omnivorous experiments. 2013 Southern Section - American Society of Plant Biologists (SS-ASPB), Little Rock, Erickson, A.A., Paul, V.J., Kwiatkowski, L.M., Becerro, M., AR: Rhizophytic green algal extracts differentially affect feeding by sea urchin species. 2012 LSUS School of Mathematics and Science Student Research Forum, Shreveport, LA: Faith, C., Chaniyara, R., Erickson, A., Do nutrient additions influence herbivory on mangrove leaves?

2011LSUS Student Research Forum, Shreveport, LA: Edwards, M.R., Lincoln, A.,Holstein,H., Erickson, A.A., Behavior of the mangrove tree crab duringits active season.

2010	LSUS Student Research Forum, Shreveport, LA: Morlock, T., Erickson, A.A., Proposed
	methods for examining the effects of nutrient fertilization on nutritional and
defens	ive chemistry and herbivore feeding behavior in mangrove forests.
2010	39th Annual Benthic Ecology Meeting, Wilmington, NC: Erickson, A.A., Feller, I.C., Paul,
	V.J., Kwiatkowski, L.M., West, L., Trejo, P., Red mangrove leaves improve with
age	like fine wine: the relationship between crab feeding, leaf age, and pre-existing
	herbivore damage.
2009	38 th Annual Benthic Ecology Meeting, Corpus Christi, TX: Erickson, A.A., Lee, W.,
	Hurricane impact on mangrove tree crab populations in Florida.
2007	29 th Annual Southeastern Phycological Colloquy, Dauphin Island, AL: Erickson,
A.A.,	Paul, V.J., Becerro, M., Duran, S., and Kwiatkowski, L.M., The role of
chemic	cal defense in the palatability of congeneric, rhizophytic, green
algae.	
2005	27 th Annual Southeastern Phycological Colloquy, Fort Pierce, FL: Erickson, A.A.,
	Paul, V.J., Van Alstyne, K.L., and Kwiatkowski, L.M., Palatability of green
	algae that employ different types of activated chemical defenses.

2005 27th Annual Southeastern Phycological Colloquy, Fort Pierce, FL: Paul,

	Valerie J., Ritson-Williams, Raphael, Erickson, Amy A., and Becerro, Mikel,
	Susceptibility of marine macroalgae and cyanobacteria to herbivorous
fishes a	and the sea urchin <i>Diadema antillarum</i> .
2005	27 th Annual Southeastern Phycological Colloquy, Fort Pierce, FL: Paul,
	Valerie J., Becerro, Mikel, Bonito, Victor, Ritson-Williams, Raphael, and
	Erickson, Amy, Impacts of benthic marine cyanobacteria on coral reefs:
	ecology and toxicity.
2005	27th Annual Southeastern Phycological Colloquy, Fort Pierce, FL: Puglisi,
	Melany, Ross, Cliff, Erickson, Amy, and Paul, Valerie, Chemical defenses of
	the marine cyanobacterium <i>Lyngbya</i> cf. <i>confervoides</i> .
2004	The Center of Excellence in Biomedical and Marine Biotechnology Meeting, Boca
	Raton, FL: Paul, Valerie J., Becerro, Mikel, Erickson, Amy, Duran, Sandra,
	and Ritson-Williams, Raphael, Recent Research from the Smithsonian Marine
	Station at Fort Pierce.
2004	Tenth International Coral Reef Symposium, Okinawa, Japan: Paul, Valerie J., Ritson-
	Williams, Raphael, Erickson, Amy A., and Becerro, Mikel,
Suscep	tibility of marine macroalgae and cyanobacteria to herbivorous
fishes a	and the sea urchin Diadema antillarum.
2004	Tenth International Coral Reef Symposium, Okinawa, Japan: Paul, Valerie J.,
	Becerro, Mikel, Bonito, Victor, Ritson-Williams, Raphael, and Erickson,
	Amy, Impacts of benthic marine cyanobacteria on coral reefs: ecology and toxicity.
2003	Twin Cays Mangrove Ecosystem, Belize: Biodiversity, Geological History and Two
	Decades of Change, Fort Pierce, FL: Erickson, Amy A., Bell, Susan S., and
	Dawes, Clinton J., Relating mangrove herbivore preference and consumption
	to leaf chemistry and structure.
2003	Estuarine Research Federation, Seattle, WA: Erickson, Amy A., Bell, Susan
	S., and Dawes, Clinton J., Are similar conclusions found when measuring
	herbivory in various ways?: Implications for understanding ecology and
	addressing management issues.
2003	Benthic Ecology Meeting, Groton, CT: Erickson, Amy A., Bell, Susan S., and Dawes,
	Clinton J., Do different measures of herbivory lead to similar
conclu	sions?
2001	Estuarine Research Federation, St. Petersburg, FL: Erickson, Amy A., Saltis,
	Mark, Bell, Susan S., and Dawes, Clinton J., Dietary preference for red
	mangrove leaves as measured by leaf damage and crab gut contents.
2001	Benthic Ecology Meeting, Durham, NH: Erickson, Amy A., Saltis, Mark, Bell, Susan
S.,	and Dawes, Clinton J., Determining diet preference for mangrove
species	-
1999	Ecological Society of America, Spokane, WA: Erickson, Amy A., Bell, Susan S.,
and	Dawes, Clinton J., Crab herbivory on the red mangrove Rhizophora
mangle	
	ck mangrove Avicennia germinans L.
1999	Benthic Ecology Meeting, Baton Rouge, LA: Erickson, Amy A., Crab herbivory in
	mixed versus pure stands of the red mangrove <i>Rhizophora mangle</i> L.

Invited talks and other presentations:

2020	UC Davis Pre-Health Conference: Panelist on Health science MS & post-bacc
	programs
2020	Salvinia working group: LSUS Giant Salvinia control research teamworking
	group; Erickson, A., Mast, J.
2019	LSUS Faculty research showcase: Control of Salvinia molesta and eDNA
2017-	Medical Seminar, LSUS: Mangroves in Medicine
2015	College of Arts and Sciences Lecture Series, LSUS: Associational defense
	protects mangroves from crab herbivory
2014	College of Arts and Sciences Lecture Series, LSUS: Palatability of green
	algae that employ different types of chemical defenses
2014	LSU Day: To the sea and back: Marine Biology at LSUS
2013	College of Arts and Sciences Lecture Series, LSUS: Sea urchin species vary
	in their susceptibility to algal chemical defenses
2013	College of Arts and Sciences Lecture Series, LSUS: To the sea and back:
	Marine Biology at LSUS
2012	Louisiana School of Mathematics, Sciences, and the Arts: Marine biology in
	Northern Louisiana?
2007	LSUS: To eat or not to eat? Chemical mediation of feeding preferences for the
	mangrove tree crab Aratus pisonii
2006	Smithsonian Marine Station Lecture Series, LSUS: Palatability of green algae
	that employ different types of chemical defenses
2006	Georgia Tech: Community Ecology
2004	Smithsonian Marine Station Lecture Series, LSUS: Relating mangrove
	herbivore preference and consumption to leaf chemistry and structure
1998	Science Day at magnet school, Tampa, FL: Mangrove crabs
1998	Highschool Biology Honors program: Mangrove crabs
Research Ex	perience:
2020-	Environmental DNA studies on marine and freshwater species, LSUS
2019-	Effects of acidification of freshwater organisms, LSUS
2019-	Invertebrate diversity in Salvinia molesta, LSUS
2018-	Effects of Salvinia molesta extracts on microbial growth, LSUS
2016-	Allelopathic effects of common aquatic plants on the invasive water fern
Salvinia	molesta, LSUS
2003-	Marine chemical ecology, feeding preference studies, chemical extraction &
	isolation, Louisiana State University Shreveport, Smithsonian Marine
	Station, University of South Florida
1999	Drift algae sampling, Dept. of Biology, University of South Florida
1998-2003	Mangrove forestry, herbivory, leaf chemistry, & gut content analysis, Dept. of
1990 2000	Biology, University of South Florida
1998	Mangrove forestry of Central and South Florida, Dept. of Biology, University
of	South Florida
1997	Seagrass mapping and prop-scar study, Dept. of Biology, University of
	South Florida
1996-1997	Morphological, distributional, & chemical studies of rhizophytic green
	macroalgae, Dept. of Biology, University of South Florida
	muerouigue, Dept. of Diology, Oniversity of South Fiorian

1994-1997	Anatomical, histological, & receptor-binding of GABAA receptors in chick
	brainstem auditory nuclei, Dept. of Psychology, Florida State
University	
1992-1994	Feeding preference studies, Dept. of Biology, Georgetown University
1992-1994	Behind the Scenes Volunteer, Smithsonian Institution: Everglades Mesocosm
	Project

Professional Affiliations:

2013-	American Society of Plant Biologists (National, Southern
Section)	
2013-2014	Botanical Society of America
2013-2017	American Chemical Society (National, LSUS)
2011-2012	Gulf Estuarine Research Society
2006-2007	Society of Wetland Scientists
2004-2007	Association of Tropical Biology and Conservation
2000-2006	Georgetown University Alumni Admissions Program
2000-2007, 2010-	Phi Kappa Phi Honor Society
2000-	Southeastern Estuarine Research Society
2000-	Coastal and Estuarine Research Federation
1999-	Ecological Society of America

Reviewer:

Acta Oecologica, African Journal of Aquatic Science, Aquatic Biology, Aquatic Botany, Aquatic Ecology, Bulletin of Marine Science, Ecology, Estuaries and Coasts, Estuarine, Coastal, and Shelf Science, EurAsian Journal of BioSciences, Florida Scientist, Hydrobiologia, Journal of Animal Ecology, Journal of the Marine Biological Association of the United Kingdom, Journal of Chemical Ecology, Marine Biology, Marine Ecology Progress Series, Nature Chemical Ecology, Oecologia, Oxford University Press Review Panel, Plos One, Regional Environmental Change, Restoration Ecology, Wetlands Ecology and Management

Service:

2021	Member, CERF Niering Award Committee
2021	Member, Chair of Biological Sciences Search Committee
2021	Member, Biological Sciences Search Committee
2020	Member, Biology Curriculum revision (with V. Salim)
2020	Member, Anatomy & Physiology Search Committee
2020	Member, Graduate Thesis Committee Compensation Taskforce
2019	Member, Graduate Manual Revision Taskforce
2019-	Member, Graduate Council
2019-2020	Member, Red River Watershed Management Institute Director Search
Committee	
2019-2020	Member, Assessment Committee
2019	Create permanent Salvinia molesta display at the Shreveport Aquarium
2018-	Member and Chair, GIS Committee
2018-2019	Member, Associate Vice Chancellor/Director of Online Learning Search
	Committee

2019	Ded Diver Netional Wildlife Defree Calebration
2018	Red River National Wildlife Refuge Celebration
2018	Minden Maker's Fair
2018	Shreveport/Bossier Maker Faire
2018	Create LSUS Salvinia molesta brochure
2018	Member, Cell Biology Search Committee
2017-	Member, Internship Committee
2017-	Member, Library Committee
2017	Member, Badge Committee
2017	Member, General Education Working Group
2016	Member, Online Teaching Policy Committee
2016-2018	Advisor, Pre-Vet Club
2016-2017	Chair, Library Committee
2016-2017	Member, LSUS Strategy Facilitation Team
2016	Member, College of Arts and Sciences Freshman Advisor Search Committee
2016	Chair, Director of Online Learning Search Committee
2015-2017	Member, Retention Committee
2015-2016	Member, Ad-hoc ADA Guidelines Committee
2014-	Advisor, Biology Club
2014-2015	Chair, Instruction and Professional Development Committee, LSUS
2014-2015	Member, Faculty Credentialing Committee
2014	Member, Grievance Committee
2014	Member, Ad-hoc Policy Committee
2013-	Chair, LSUS School of Mathematics and Science Scholarship Committee
2013-2017	LSUS American Chemical Society Chapter
2013-2014	Member, LSUS SACS-COC Reaffirmation Committee
2013	Member, 8-week Session Faculty Focus Group
2012-2014	Creator & Manager, LSUS Ecology & Environmental Science Advising Page
2012-2014	Secretary, Faculty Senate Executive Committee, LSUS Faculty Senate
2012	Evaluator, NSF-funded Center for Ocean Science Education Excellence
(COSEE)	Scientist Case Studies
2012	Member, Biology Curriculum Review Committee
2012	Member, Ad-hoc Senate Committee on Revenue Generation
2012	Member, Senate Ad-hoc Committee: CLEP tests
2012	Senator at Large, Senate Executive Committee, LSUS Faculty Senate
2011-2015	Advisor, SOAR Advising to Incoming Freshman
2011-2013	Senator, LSUS Faculty Senate
2011-2013	Member, LSUS Faculty Progress Report Review Committee
2011-2013	Member, LSUS Assessment Committee
2011-2012	Judge, Senior Projects at Airline HS
2011	Member, Senate Ad-hoc Committee: Development of a Promise Program
2009-2017	Member, Distance Learning Council
2009-2017	Member, M.S. in Biological Sciences Applicant Review Committee, Dept. of
2009	Biological Sciences, LSUS
2008-2012	Member, LSUS School of Science Scholarship Committee
2008-2012	Coordinator, Shreveport-Bossier SCI-NET Research Experiences Post Katrina
	grant

2008-2012	Member, Post-Katrina S/B Sci-Net Committee
2008-2012	Member, M.S. in Environmental Biology Development Committee, Dept. of
	Biological Sciences, LSUS
2008	Member, Introductory Biology Curriculum Committee, Dept. of Biological
	Sciences, LSUS
2007-	Biological Sciences Advisor, LSUS
2007-2013	Member, Instruction and Professional Development Committee, LSUS
2000-2006	Interviewer, Georgetown University Alumni Admissions Program
1998-1999	Undergraduate Advisor, Biology and Pre-Professional Sciences, USF

Thesis students:

Advisor:

High school: Jonathan Woldie, 2019-2020 Elijah Burks, 2019-2020 Sarah Glynn, 2007

Undergraduate: Mariah Bertrand, 2019-2020 (not completed) Lauren Henderson, 2019-2020

Masters: Emily Dean, 2021-Mariah Bertrand, 2020-Erin Oostenburg, 2019-Katelyn Toms, 2018-Shana Mullins, 2018-2019 (not completed) Jessica Mast, 2016-

Committee member:

Camila Muller, 2020-Wendy Dobbins, 2018 Dominique Washington, 2014 Kyle Koho, 2007

Undergraduate research mentoring:

1999- Training undergraduate assistants (>150), Biology Dept., USF, Smithsonian Marine Station, LSUS in field and laboratory techniques for mangrove forestry, chemistry, and herbivory

Mike Maguigan



Director of the Red River Watershed Management Institute

My training is as a physical geographer, with interests in the environmental system – rock, soil, water, air, and vegetation. I also have experience in using technologies to monitor these systems, such as GIS and remote sensing. My research is a combination of field work, laboratory analyses, and computer analyses. Currently, I am interested in studying water quality and sediment transportation within the Red River Watershed, though I am open to new project to which I can lend my expertise. Questions I'm interested in answering:

- How do sediment flow and water quality vary by seasonal flow levels across the Red River Watershed?
- Can we isolate a potential cause of these phenomena at certain points in the river?
- How can technology better assist us in addressing these questions?

Elahe Mahdavian, PhD

Professor of Biochemistry PhD: University of South Carolina Office: SC312; Tel: 318-797-5227; Fax: 318-797-5090 Email: elahe.mahdavian@lsus.edu

Appointments:

- 2015-present Professor of Biochemistry, LSUS, Shreveport, LA.
- 2009-2015 Associate Professor of Biochemistry, LSUS, Shreveport, LA.
- 2003-2009 Assistant Professor of Biochemistry, LSUS, Shreveport, LA.
- 1999-2003 Assistant Professor of Chemistry, SCSU, Orangeburg, SC.

Professional Memberships & Contributions

- 2005-Present Member, American Chemical Society (ACS)
- 2007-2013 Member, American Association of Cancer Research (AACR)
- 2009-Present Member, LSUS Health Sciences Advisory Committee (HSAC)
- 2015-Present Chair, Northwest Louisiana Section of ACS (NWLA-ACS)
- 2010-present Advisor, Chemistry Club, LSUS
- 2020-present Advisor, Science Matters, LSUS

Teaching Interests (Undergraduate & Graduate Levels):

I. Biochemistry Lecture & Laboratory Courses (BCHM410/BCHM610; BCHM412/BCHM612)

- II. Computational Chemistry/Biology -CADD Courses (CHEM495/CHEM795; BIOS495/BIOS795)
- III. Research and Thesis Courses (CHEM290, CHEM490, CHEM790, BIOS790)

IV. Practical Bioinformatics Course, Team-Taught & Cross-Institutional LSUHSC (IDSP 204A/B)

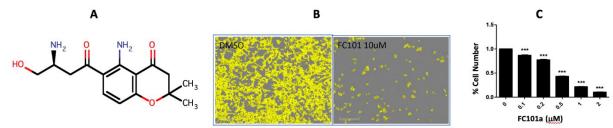
V. Chemistry Literature Course (CHEM301)

Research Interests:

My scholarly activities focus on the following two collaborative, student-centered and interdisciplinary research programs:

Cancer Research Program: This is an interdisciplinary research program spanning the interfaces of chemical synthesis, medicinal chemistry, and cancer biology fields. It is centered on a compound known as fusarochromanone (FC101), an anti-cancer agent with unique structure and function. FC101 is a cancer-specific cytotoxic agent that exhibits 10-100 times the effects against cancer cells vs. normal cells of the same type. This selectivity is the result

of differential uptake by cancer cells and increased toxicity against them. FC101 significantly inhibits proliferation/migration and induces apoptosis in cultured human cancer cells. The more invasive cancer cells, particularly the oncogenic BRAF mutant- MAPK driven cancer (e.g. melanoma, bladder, pancreatic, and TNBC), are more sensitive to the anti-cancer effect of FC101. The total synthesis of FC101 in its enantiomerically pure form was accomplished using a seven-step reaction scheme in 2014. This allowed for further investigation of its therapeutic potential and provided new opportunities to modify the molecule for lead optimization and analysis of quantitative structure activity relationships (QSAR). While FC101's exact mechanism of action is currently unknown, we have shown that it simultaneously inhibits the activity of two major oncogenic pathways, MAPK (corresponding to p-ERK reduction) and mTOR (corresponding to p-S6K, and p-S6 reduction) in cultured cancer cells. The current FC101 research focuses on: I. lead-optimization/SAR through synthesis and biological evaluation of key structural analogs; II. Computational and *in-vitro* drug screens to develop the phenotypic drug response/sensitivity profiles; III. Mechanistic understanding of how FC101 induces apoptosis and inhibits proliferation/migration in TNBC. Knowledge from computational, in-vitro, and mechanistic studies reveals FC101's molecular mechanism of action and ensures progress towards the pre-clinical stage of this research involving animal models of cancer.



A. The Chemical Structure of FC101; B. Images of FC101- and DMSO-treated cultured TNBC; C. FC101's growth inhibition of cultured TNBC.

Computer Aided Drug Discovery (CADD) Program:

This is a new interdisciplinary research/pedagogy program within the Departments of Chemistry and Biological Sciences involving CADD methodology. This approach has been incorporated into several cancer projects and new research units on antiviral therapeutics for COVID-19, practicing highly relevant science during the pandemic. Virtual screening methods, the search for bioactive compounds via computational tools, provide shortcuts and a wide range of opportunities to speed up drug discovery, while reducing the associated financial and attrition risks. The CADD methodology has also been integrated into a research-based online course, merging guided research with interactive classroom instructions. The course enables students to use researchbased methods and employ active learning with publicly available bioinformatics/structural biology data and modern computational modeling tools to identify promising anti-viral drugs for COVID-19. The inspiration for this online course is four-fold: (1) The importance of teaching science as science is practiced, merging guided research with course-based instruction to broaden student participation in research. (2) The recognition that interdisciplinary research skills in applied bioinformatics, computational modeling, and structural biology are indispensable to a student's scientific education (3) The significant negative impact of COVID-19 pandemic on public health and hence the emergent unmet need for new antiviral drugs. (4) Instructional shifts in response to COVID-19 pandemic and its impact upon the classroom-based student research experience. The CADD course, which has now been successfully implemented four times since summer 2020, combines three modules: lectures/discussions including live demos, inquiry-based assignments, and science communication.

Selected Publication:

1. Mahdavian E, Spencer HT, Dunlap RB. Kinetic Studies on Drug Resistant Variants of Escherichia coli Thymidylate Synthase: Functional Effects of Amino Acid Substitutions at Residue 4. *Arch Biochem Biophys*. **1999**; 368(2):257-64. PMID: 10441376.

2. Phan J, Mahdavian E, Nivens MC, Minor W, Berger S, Spencer HT. Catalytic Cysteine of Thymidylate Synthase is Activated Upon Substrate Binding. *Biochemistry*. **2000**; 39(23):6969-78. PMID: 10841779.

3. Tomic-Vatic A, Eytina J, Chapman J, Mahdavian E, Neuzil J, Salvatore BA. Vitamin E amides, a new class of vitamin E analogues with enhanced pro-apoptotic activity. *Int J Cancer*. **2005**; 117(2):188-93. PMID: 15900584.

4. Mahdavian E, Sangsura S, Landry G, Eytina J, Salvatore BA. A Novel Synthesis of Tocopheryl Amines and Amides. *Tetrahedron Letters*. **2009**; 50:19-21.

5. Turánek J, Wang XF, Knötigová P, Koudelka, S, Mahdavian E, Procházka L, Sangsura S, Vacek A, Salvatore BA, Neuzil J. Liposomal formulation of vitamin E analogs as an efficient and selective anti-cancer treatment" *Applied Toxicology and Pharmacology*, **2009**; 237.

6. Dong LF, Jameson VJ, Tilley D, Prochazka L, Rohlena J, Valis K, Truksa J, Zobalova R, Mahdavian E, et al. Mitochondrial Targeting of a-Tocopheryl Succinate Enhances Its Pro-

apoptotic and Anti-cancer Efficacy. *Free Radic Biol Med*. **2011**; 50(11):1546-55. PMID: 21402148.

7. Dong L, Jameson VJ, Tilly D, Cerny J, Mahdavian E, Marín-Hernández A, et al. Mitochondrial Targeting of Vitamin E Succinate Enhances Transmission of its Activity via Mitochondrial Complex II. *J Biol Chem*. **2011**; 286(5):3717-28. PMID: 21059645.

8. Ying G, Chen X, Chang C, Singh K, Barzegar M, Mahdavian E, et al. "Fusarochromanone Induces G1 Cell Cycle Arrest and Apoptosis in COS7 and HEK293 Cells. PLOS ONE. **2014**; 10(9):11. PMID: 25384025, PMCID: PMC4226581.

9. Mahdavian E, Williams-Hart T, Furmanski B, Kevil C, Gopi K, Yoon-Jee K, et al. "Biological Activities of Fusarochromanone: A Potent Anti-Cancer Agent". *BMC Research Notes*. **2014**; 7:601. PMID: 25187308, PMCID: PMC4168212.

10. Mahdavian E, Marshall M, Martin P, Cagle P, Salvatore BA, Quick Q. "Caspase Dependent Signaling Underlies Glioblastoma Cell Death in Response to the Fungal Metabolite, Fusarochromanone". *Int J Mol Med*. **2014**; 34(3):880-5. PMID: 25016928, PMCID: PMC4121350.

11. Wynne M, Salvatore BA, Mahdavian E. Use of In-silico Assays for ADMET and Target Profiling of Fusarochromanone. *Journal of In-silico Pharmacology*. **2015**; 3(6). PMCID: PMC4464579.

12. Hasanain, G., Mahdavian E, et al. "Development and characterization of folic acidfunctionalized apoferritin as a delivery vehicle for epirubicin against MCF-7 breast cancer cells", *ARTIFICIAL CELLS, NANOMEDICINE, AND BIOTECHNOLOGY*, **2018**; 46, 847-854.

13. Hashempour, S., Mahdavian E, et al. "Binding Studies of AICAR and Human Serum Albumin by Spectroscopic, Theoretical, and Computational Methods" *Molecules*, 2020; 25(22).

14. Kashanian, S., Mahdavian E, et al. "Novel dual targeting system based on cSLNs and modified apoferritin to simultaneously deliver doxorubicin and mitoxantrone anticancer drugs" *Current Pharmaceutical Biotechnology*, **2021**; 26(6).

15. Burford, N.; Smith, A.; Salvatore, B.; Mahdavian, E.; "Combination Therapies for TNBC: EGFR inhibitors and Fusarochromanone". *Journal of BMC Cancer*, **2021**. In Revision.

16. Mackay, R.; Weinberger, P.; Copland, J.; Mahdavian, E.; Xu' Q.; "YM155 induces DNA damage and cell death in anaplastic thyroid cancer cells by inhibiting DNA topoisomerase $II\alpha$ ".

Molecular Oncology (ID: MOLONC-21-0535). 2021. Submitted.

17. Salvatore, B., Mahdavian, E., "Phenotypic Investigations of Combination Therapeutics for Metastatic Cancer". Mini Review, *BMC Medicine*. **2021**. In Prep.

List of published work in NCBI- My Bibliography http://www.ncbi.nlm.nih.gov/pmc/?term=elahe+mahdavian Google Scholar Website: <u>https://rb.gy/eerhtg</u>

Stuart Nielsen, PhD



Title: Assistant Professor Start Year at LSUS: 2021 Department: Biological Sciences Department School: Mathematics and Sciences College: Arts and Sciences Office Location: Science Building 220 Office Phone: 318-797-2413 Office Fax: 318-797-5090 Email: stuart.nielsen@lsus.edu Office Hours: Fall 2021: M 12:00-14:00; T/R 13:00-17:00 (or virtually via Zoom).

Teaching Assignments:

BIOS 101	General Biology (for non-science majors)	Fall
BIOS 109	Animal Diversity (for non-science majors)	Fall
BIOS 222L	General Zoology Laboratory	Fall & Spring
BIOS 365	Herpetology	Spring

*Please feel free to contact me for class itineraries, text books, or syllabi.

Research Interests:

I am an Evolutionary Biologist and a Herpetologist (someone that studies reptiles and amphibians).

Student Research Opportunities:

1) Molecular systematics, biogeography, and evolution of reptiles and amphibians.

One major concentration of my past research has been devoted to generating species-level phylogenies for numerous taxonomic groups, in order to provide an evolutionary framework for subsequent research. A well-resolved, well-supported phylogeny can clarify evolutionary patterns that may have been confused by a non-evolutionary taxonomy and is vital to identify evolutionary patterns. Phylogenies are an essential tool for the investigation of spatial and historical patterns of species richness, and the influence of environment and geography on generating and maintaining biological

diversity. I have used time-calibrated molecular phylogenies to study past geoclimatic change on speciation and diversification in Africa, Australia, and New Zealand, extinction risk of range-restricted taxa, as well as the evolution of novel anti-predator defenses.

Research at LSUS will focus on three broad topics: 1) completing ongoing and initiating new projects to test complex bio-/phylogeographic hypotheses and delimit species using both phylogenomic and multi-locus coalescent methods (African lizards and frogs; Caribbean/Middle American sphaerodactylid geckos); 2) examining the origin and maintenance of species boundaries in relation to the genomic influence of sex chromosomes (many squamate lineages; see below); and 3) studying one-off evolutionary novelties including i) a unique, anti-predator, 'deimatic' startle display that has convergently evolved twice within diplodactylid geckos in Australia and New Caledonia (check out https://youtu.be/idipffPIGNg) and ii) body size extremes including both dwarfism (sphaerodactylid geckos) and gigantism (diplodactylid geckos). Data collection and analysis for most of these projects is relatively straightforward, making them ideal student research projects.

2) Sex chromosome evolution in squamates (lizards and snakes).

I am working towards understanding fundamental questions regarding the evolution, structure, and function of sex chromosomes and sex determination, and the development of new model clades to address these questions. Sex chromosomes play a central role in development by controlling sex determination, the mechanism that decides the sex of an embryo. Sex chromosomes also exert tremendous influence over essential evolutionary processes, such as speciation, sex-specific adaptation, and genetic conflict. Squamate reptiles (~10,000 species of lizards, snakes, and amphisbaenians) are ideally suited to test hypotheses about the origins and evolution of sex chromosomes because they exhibit myriad sex determining modes, including temperature-dependent (TSD) and genetic (GSD) sex determination, with both male (XX/XY) and female (ZZ/ZW) heterogamety, and many independent transitions among them. Yet, surprisingly, even at the family level, we don't know the sex chromosome systems in the vast majority (~80%) of squamate lineages. Developing new model clades is vital to advance our understanding of sex chromosomes and sex determination, beyond the limited knowledge obtained by only studying a handful of well-studied model organisms.

Student research opportunities involve continuing to identify sex chromosomes in understudied clades, particularly those of conservation concern, and assessing homology via the sequencing and assembly of genomes/transcriptomes and advanced cytogenetics.

Most Recent Publications:

- Rovatsos, M., T. Gamble, S.V. Nielsen, A. Georges, T. Ezaz, & L. Kratochvíl. 2021. Do male and female heterogamety really differ in expression regulation? Lack of global dosage balance in pygopodid geckos. Philosophical Transactions of the Royal Society B 376: 20200102. https://doi.org/10.1098/rstb.2020.0102
- Paluh, D.J., K. Riddell, C.M. Early, M.M. Hantak, G.F.M. Jongsma, R.M. Keeffe, F.M. Silva, S.V. Nielsen, M.C. Vallejo-Pareja, E.L. Stanley, D.C. Blackburn. 2021. Rampant tooth loss

across 200 million years of frog evolution. eLife 10:e66926. https://doi.org/10.7554/eLife.66926

- Knox, C., R. Hitchmough, S.V. Nielsen, T. Jewell, & T. Bell. 2021. A new, enigmatic species of black-eyed gecko (Reptilia: Diplodactylidae: Mokopirirakau) from North Otago, New Zealand. Zootaxa 4964(1):140–156. https://doi.org/10.11646/zootaxa.4964.1.7
- Heinicke, M.P., M.H. Beidoun, S.V. Nielsen, & A.M. Bauer. 2021. Phylogenetic analysis of "Breviceps adspersus" documents B. passmorei Minter et al., 2017 in Limpopo Province, South Africa. Herpetology Notes 14:397–406.
- Hitchmough, R.A., S.V. Nielsen, J.A. Lysaght, & A.M. Bauer. 2021. A new species of Naultinus from the Te Paki area, northern New Zealand. Zootaxa 4915(3):389–400. https://doi.org/10.11646/zootaxa.4915.3.7
- Keating, S.E., M. Blumer, L.L. Grismer, A. Lin, S.V. Nielsen, M.K. Thura, P.L. Wood Jr., E.S.H. Quah, & T. Gamble. 2021. Sex chromosome turnover in bent-toed geckos (Cyrtodactylus). Genes 12:116. https://doi.org/10.3390/genes12010116

Academic Positions Held:

Assistant Professor

• Department of Biology, Louisiana State University in Shreveport, Shreveport, LA, USA (2021-present)

Courtesy Faculty

• Division of Herpetology, Department of Natural History, Florida Museum of Natural History, University of Florida, Gainesville, FL (2021-present)

Adjunct Professor/Visiting Assistant Professor

- Sciences for Health Programs Department, Santa Fe College, Gainesville, FL (2021-2021)
- Department of Biological Sciences, Marquette University, Milwaukee, WI (2018-2019)

Postdoctoral Researcher

- Department of Natural Sciences, University of Michigan–Dearborn, Dearborn, MI. Advisor: Dr. Matt Heinicke (2020-2020)
- Division of Herpetology, Florida Museum of Natural History, University of Florida, Gainesville, FL. Advisor: Dr. Dave Blackburn (2018-2020)
- Department of Biological Sciences, Marquette University, Milwaukee, WI. Advisor: Dr. Tony Gamble (2016-2018)

Molecular Lab Manager

- Division of Herpetology, Florida Museum of Natural History, University of Florida. (2018-2020)
- Department of Biological Sciences, Marquette University, Milwaukee, WI. (2016-2018)

Post-PhD Fellowships Awarded

- US Dept. of State, J. William Fulbright Postdoctoral Reseach Fellowship (Brazil): Instituto Nacional de Pesquisas da Amazônia, Manaus, Brazil. Advisor: Dr. Fernanda Werneck (Sex chromosome evolution in geckos) (Delayed due to COVID-19)
- John J. Brander and Christine E. Rundblad Research Fellowship: Milwaukee Public Museum, Milwaukee, WI. (Declined)
- Endeavour Postdoctoral Fellowship: Australian National University, Canberra, Australia. Advisors: Drs. Paul Oliver, Scott Keogh, & Craig Moritz (High turnover in sex chromosomes within Australia's Gehyra geckos; New Guinea gekkonid taxonomy) (2016–2017)

Vonny Salim, PhD



Title: Assistant Professor Start Year at LSUS: 2018 **Department:** Biological Sciences School: Mathematics and Sciences **College:** Arts and Sciences Office Location: SC 212 Office Phone: 318-797-5228 Office Fax: 318-797-5222 Email: Vonny.Salim@lsus.edu **Current Courses Taught:** BIOS 420 Cell Biology Fall Cell Biology (Graduate Level) **BIOS 620** Fall Cell Biology Laboratory Fall BIOS 420L BIOS 620L Cell Biology Laboratory (Graduate Level) Fall Spring BIOS 430 Molecular Biology Molecular Biology (Graduate Level) **BIOS 630** Spring BIOS 430L Molecular Biology Laboratory Spring BIOS 630L Molecular Biology Laboratory (Graduate Level) Spring BIOS 351 Medical Microbiology Spring/Summer BIOS 465 Applied Biotechnology Spring BIOS 665 Applied Biotechnology (Graduate Level) Spring **BIOS 107** Humans and Their Environment Summer BIOS 490 Cancer Biology Summer Cancer Biology (Graduate Level) BIOS 690 Summer **Undergraduate Research** BIOS 491 Fall

Research Interests:

My research interests are in the biosynthesis, function, and application of plant-derived natural products for human health and the cellular machineries and dynamics of phytochemicals for further integration into human disease treatment and prevention, such as in cancer therapy.

 Genomics and metabolomics of natural product biosynthesis for human health. Medicinal plants produce a wide variety of chemicals with pharmaceutical values. One class of plant specialized metabolites, alkaloids have been used as chemotherapeutic agents. Alkaloids, such as vinblastine harvested from *Catharanthus roseus* and camptothecin from *Camptotheca acuminata* have been used in the clinic, however, their productions are still limited by expensive and tedious isolation processes from plants. The goal of this project to elucidate anticancer alkaloid biosynthetic pathways and functionally characterize enzymes involved in the production of these high-value compounds. Large-scale sequencing of medicinal plants that produce alkaloids have accelerated the identification process of putative biosynthetic genes. In this project, we investigate the aspects of genomics and metabolomics of natural product biosynthesis and metabolic engineering to increase the production of alkaloids and other phytochemicals with important biological activities for human health.

2. Metabolite and enzyme engineering of natural product biosynthetic pathways in microbial systems.

Recombinant expression of plant biosynthetic genes in microbial systems, such as bacteria *Escherichia coli* and *Saccharomyces cerevisiae* (yeast) has been known as an efficient method to investigate the function of novel biosynthetic genes. In our laboratory, we utilize reverse genetics approaches, including virus-induced gene silencing and RNA interference to test the function of putative genes involved in the biosynthesis of alkaloids and other phytochemicals. Further genetic manipulations of plants and microbial systems in our laboratory also include overexpression and mutagenesis that may result in accumulation of novel compounds with potential biological activities. We aim to generate chemical diversity of lead compounds, especially those with anticancer properties and accelerate drug discovery.

3. Functional characterization of microbiomes in medicinal plant systems for applications and impacts on human health.

Alkaloids are well-known targets in anticancer drug development. The ability of medicinal plants to produce cytotoxic alkaloids involves biotic factors, such as their relationships with microbes and utilization of natural product secretion systems. In this project, microbial communities associated with the production of anticancer alkaloids are identified using Next Generation Sequencing (NGS) machine, and we also perform the metabolite analysis using Ultra-Performance Liquid Chromatography-Mass Spectrometry (UPLC-MS) located at the LSUS Cyber Collaboratory. The integration of metagenomics and metabolomics accelerates the efforts to determine specific microbial colonization that modulate the anticancer alkaloid metabolism in medicinal plants.

List of publications:

Nasirian, V., Shamsipur, M., Molaabasi F., Mansouri, K., Sarparast, M., **Salim, V**., Barati, A., Kashanian, S. (2020). miRNA-21 rapid diagnosis by one-pot synthesis of highly luminescent red emissive silver nanoclusters/DNA. *Sensors and Actuators B: Chemical* 308: 127673.

Salim, V., Jones, A.D., DellaPenna, D. (2018). *Camptotheca acuminata* 10-hydroxycamptothecin *O*-methyltransferase: An alkaloid biosynthetic enzyme coopted from flavonoid metabolism. *Plant Journal* 95:112-125.

Qu, Y., Easson, M., Simionescu, R., Hajicek, J., Thamm, A.M.K., **Salim, V.**, De Luca, V. (2018). Solution of the multistep pathway for assembly of corynanthean, strychnos, iboga and aspidosperma monoterpenoid indole alkaloids from 19*E*-geissoschizine. *PNAS* 115: 3180-3185.

Sadre, R., Magallanes-Lundback, M., Pradhan, S., **Salim, V**., Mesberg, A., Jones, A.D., DellaPenna, D. (2016). Metabolite diversity in alkaloid biosynthesis: A multi-lane (diastereomer) highway for camptothecin synthesis in *Camptotheca acuminata*. *Plant Cell* 28:1926-1944.

De Luca, V., **Salim, V**., Thamm, A., Masada-Atsumi, S., Yu, F. (2014). Making iridoids/secoiridoids and monoterpenoid indole alkaloids: Progress on pathway elucidation. *Current Opinion in Plant Biology* 19:35-42.

Salim, V., Wiens, B., Masada-Atsumi, S., Yu, F., De Luca, V. (2014). Iridodial oxidase catalyzes a key 3 step oxidation to form 7-deoxyloganetic acid in the *Catharanthus roseus* iridoid biosynthesis. *Phytochemistry* 101:23-31.

Besseau, S., Kellner, F., Lanoue, A., Thamm, A.M.K., **Salim, V**., Schneider, B., Geu-Flores, F., Höfer, R., Guirimand, G., Guihur, A., Oudin, A., Glevarec, G., Foureau, E., Papon, N., Clastre, M., Giglioli-Guivarc'h, N., St-Pierre, B., Werck-Reichhart, D., Burlat, V., De Luca, V., O'Connor, S.E., Courdavault, V. (2013). A pair of tabersonine 16-hydroxylases initiates the synthesis of vindoline in an organ-dependent manner in *Catharanthus roseus*. *Plant Physiology* 163: 1792-1803.

Salim, V., Yu, F., Altarejos, J., De Luca, V. (2013). Virus-induced gene silencing identifies *Catharanthus roseus* 7-deoxyloganic acid-7-hydroxylase, a step in iridoid and monoterpene indole alkaloid biosynthesis. *Plant Journal* 76: 754-765.

Asada, K.*, **Salim, V**.*, Masada-Atsumi, S.*, Edmunds, E., Nagatoshi, M., Terasaka, K., Mizukami, H., De Luca, V. (2013). A 7-deoxyloganetic acid glucosyltransferase contributes a key step in secologanin biosynthesis in Madagascar periwinkle. *Plant Cell* 25: 4123-4134. (* co-1st authors).

Salim, V., De Luca, V. (2013). Towards complete elucidation of monoterpene indole alkaloid biosynthesis pathway: *Catharanthus roseus* as a pioneer system. *Advances in Botanical Research* 68: 1-37.

De Luca, V., **Salim, V**., Levac, D., Atsumi, S. M., Yu, F. (2012). Discovery and functional analysis of monoterpenoid indole alkaloid pathways in plants. *Methods in Enzymology* 515: 207-229.

De Luca, V., **Salim, V**., Atsumi, S.M., Yu, F. (2012). Mining the biodiversity of plants: A revolution in the making. *Science* 336: 1658-1661.

Roepke, J.*, **Salim, V**.*, Wu, M., Thamm, A.M.K., Murata, J., Ploss, K., Boland, W., De Luca, V. (2010) Vinca drug components accumulate exclusively in leaf exudates of Madagascar periwinkle. *PNAS* 107: 15287-15292. (* co-1st authors).

Biography:

Postdoctoral position:

Research Associate, Michigan State University, Lansing, Michigan, Department of Biochemistry and Molecular Biology, Supervisor: Dr. Dean DellaPenna, 2013-2018.

Education:

Doctor of Philosophy in Biotechnology, Brock University, St. Catharines, Ontario, Canada, Supervisor: Dr. Vincenzo De Luca, 2013.

Bachelor of Science in Biotechnology/Biochemistry, Calvin University, Grand Rapids, Michigan, 2007.

Professional Experience:

Development Assistant, Alder Biopharmaceuticals, Inc., Bothell, Washington, 2007-2008.

Quality Assurance Assistant, MDS Pharma Services, Bothell, Washington, 2007. Brain Tumor Research Data Assistant, New Approaches to Brain Tumor Therapy (NABTT), the Sidney Kimmel Comprehensive Cancer Center at Johns Hopkins Hospital, Baltimore, Maryland, 2005.

Peter Siska, PhD

Professor of Biological Sciences and Geosciences Education: RNDr. Komensky University, Bratislava Ph.D. Komensky University, Bratislava, Slovakia Ph.D Texas A&M University, TX, USA

Courses: BIOS 110 Principles of Biology BIOS 210 Introduction to Biological Inquiry ENSC 200 Introduction to geographic information systems ENSC 300 Introduction to remote sensing ENSC 300 Intermediate geographic Information system BIOS 490 Special Topics in Biological Sciences BIOS 490 Seminar ENSC 690

Research interest:

1. Palynology, ecosystems and environment

Pollen grains are one of the most interesting elements of biosystem. They are responsible for transfer of genetic information and health of ecosystems; the fossil pollen grains are also indicators of past climate change. In addition, pollen grains are used in forensic science to trace criminal activities including illegal business with honey.

2. Geographic information systems to natural resource management studies

Investigating spatial patterns of natural and human phenomena such as radioactivity and its impact on human health; investigating spatial pattern of toxic elements and their impact on biosphere; evaluating total stem volume using terrain, satellite data and sequential gaussian simulation.

3. Application of geostatistics in prediction of karst hazards.

The methods of kriging interpolation and variogram modeling became widely used in natural sciences. The karst environment is prone to sudden subsidence of dolines that cause economical damages and pose also danger to human life. The evaluations of parameters that lead to collapse of dolines using kriging interpolation methods can predict the potential hazards in carbonatic rock complexes.

4. Multifactorial statistics and regional dynamics in natural ecosystems

The natural ecosystems are complex entities consisting from biotic and abiotic components. Multifactorial analysis can be used to evaluate causes of dynamic changes in ecosystems and identify relatively homogeneous complexes that can be delineated and understood with respect of their internal structure and interregional dynamics.

Examples of peer reviewed publications:

Siska, P. P. Bryant, V.M. Hung, I-Kuai and P. Goovaerts (2019). Matching the Spatial Distribution of Upland and Lowland Pollen Grains with the Temperature and Humidity in the Columbia Basin. Athens Journal of Sciences, Volume 6 (1). Pgs. 1 – 18.

https://www.athensjournals.gr/sciences/2019-6-1-1-Siska.pdf

Siska, P. and I-K Hung (2017). Land Management decisions in a carbonatic geoenvironment. *IOP Conference Series: Earth and Environmental Science, Volume 92.*

Siska, P.P., Goovaerts P. and I-K Hung (2016). Evaluating susceptibility of dolines (sinkholes) for collapse in Sango, Tennessee, USA. Progress in Physical Geography DOI: 10.1177/0309133316638816.

Siska P. and S. Polacik (2014). Influence of environmental parameters on spatial distribution of pollen

grains in Columbia Basin. *Proceedings of the 2014 International Conference on Power* Systems, Energy, Environment. Interlaken, Switzerland. Pgs.116-125.

Watson, J. R., Siska, P.P. and R. L. Wolfel (2013). Assessing Gains in Language Proficiency, Cross-Cultural Competence and Regional Awareness during Study Abroad: A Preliminary Study. *Foreign Language Annals* Vol. 46 (1), pp 62-79.

Siska, P. P, I.K. Hung and V.M. Bryant. (2012). The Mapping Composite Pollen from Point Sampled Data and Cartographic Generalization. Papers of Applied Geography Conferences, Volume 35(2012): 192-201.

Siska, P. P. and V. Lauko (2011). Strategic Missile Deployment: A Geographic Perspective. Applied Geography. Elsevier Science. *Applied Geography* Volume (31): 829-838.

Siska, P.P. and L.J. Hummel (2011). The Regional Knowledge System: A Complex Response to Complex Conflicts. American Intelligence Journal. Vol 29, No: 2. 107-115.

Siska, P. P. (2009). The Deployment of Ground Based Mid-Course Missiles in Central Europe: Regional Perception and Viability. *Papers of the Applied Geography Conferences.* Volume 32 (2009): 105-114.

Kemmerly, P.R. and Peter P. Siska (2008). Karst Modeling and Hazard Assessment on the Pennyroyal Plain and Western Highland Rim. *Journal of Geography (Geografický Časopis).* 60(3): 217-240.

Siska, P. P. and P. R. Kemmerly (2008). Doline Feature Database (DFD) Development and Assessment of Hazards Using Co-Regionalization Model. In: Papers of the Applied Geography Conferences. Volume (31): 34-42.

Siska, P.P. Bryant, V.M. and I-Kuai Hung (2006). Determining Spatial Correlation in Distribution of Southern Pine Biome and Dispersion of Pine Pollen Using Geographic Information Science. *Journal of Geography (Geograficky Casopis)*. 58 (4): 239 – 258.

Siska, P. P. Goovaerts, P., Hung, I-Kuai, and V. M. Bryant (2005). Predicting the Ordinary Kriging Errors Caused by Surface Roughness and Dissectivity. *Earth Surface Processes and Landforms*. 30(6): 601 – 612.

Siska, P. P. and V. M. Bryant Jr. (2005) Digital Ecosystem Analysis. *Papers of the Applied Geography Conferences.* Volume (28): 32-139.

Siska, P. P. (2004) A Multivariate Spatial Model for Determining Urban Development Sites in Flood Prone Coastal Areas. *Papers of the Applied Geography Conferences.* Vol 27: 75 - 83.

Erikson, M. and P.P **Siska (2003).** Replay to Marcotte's Comments on Understanding Anisotropy Computations. *Journal of Mathematical Geology.* 35(5) 683 – 700.

Siska, P. P. Nelson, R. and A. Bhowmick (2003) Lidar Data in Geographic InformationSystems: Component Object Modeling (COM) Approach. Papers of Applied Geography Conferences. Volume (26): 242-248.

Siska, P. P. Bryant, M. V. and J. Jones (2001). Spatial Modeling of Modern Pollen Rain inBig Bend National Park. Palynology: Journal of American Society of Stratigraphic Palynologists. 25(2001): 199-216.

Siska, J. J. Hurburgh, C. R. and P. P. Siska (2001). The Impact of Instrument EngineeringParameters on Spectral Reproducibility Across Filter Instruments. Journal of Near Infrared Spectroscopy Volume (9): 97-105.

Siska, P. P., and I – Kuai Hung, (2001) Progression of Errors in Applied Spatial Analysis.Papers and Proceedings of Applied Geography Conferences. Vol. 24(2001): 284-290.

Stephanie Villalba, PhD

Stephanie Vanhoof Villalba, Ph.D. Assistant Professor of Biological Sciences Phone: (318) 797-5157 E-mail: stephanie.villalba@lsus.edu

Education

- 2012-2017 Louisiana State University Health Sciences Center, Shreveport, LA
 Ph.D., Cellular Biology and Anatomy
 Mentor, Edward Glasscock, Ph.D., Associate Professor
 Dissertation title: "The Role of Pharmacogenetics in the Treatment of
 Neurocardiac Dysfunction in Two Mouse Models of Epilepsy"
- 2005-2007 Louisiana Tech University, Ruston, LA M.S., Biology
- 1999-2002 Louisiana Tech University, Ruston, LA B.S., Cell and Molecular Biology Minor, Chemistry Summa cum laude

Teaching and Research Positions

2020- Assistant Professor, Department of Biological Sciences Louisiana State University Shreveport, Shreveport, LA

2020- Gratis Assistant Professor, Department of Molecular and Cellular Physiology Louisiana State University Health Sciences Center, Shreveport, LA

2017-2020 Instructor, Department of Cellular Biology and Anatomy Louisiana State University Health Sciences Center, Shreveport, LA

2012-2017 Graduate Assistant, Department of Cellular Biology and Anatomy Louisiana State University Health Sciences Center, Shreveport, LA Ph.D. Dissertation Lab of Edward Glasscock, Ph.D.

2012-2017 Graduate Teaching Assistant, Department of Cellular Biology and Anatomy Louisiana State University Health Sciences Center, Shreveport, LA

2008-2012 Instructor, Department of Biology Grambling State University, Grambling, LA

2008 Research Assistant, Department of Basic Pharmaceutical Sciences University of Louisiana at Monroe, Monroe, LA Lab of Seetharama Satyanarayanajois, Ph.D.

2006	Graduate Assistant, College of Engineering & Science Louisiana Tech University, Ruston, LA Lab of David K. Mills, Ph.D.
2005-2007	Teaching Assistant, Department of Biological Sciences Louisiana Tech University, Ruston, LA
2005-2007	Graduate Assistant, Department of Biological Sciences Louisiana Tech University, Ruston, LA Lab of David K. Mills, Ph.D.
2001-2002	Undergraduate Research Assistant, Department of Biological Sciences Louisiana Tech University, Ruston, LA Lab of Wendy Trzyna, Ph.D.
2001	Summer Undergraduate Research Fellow, Department of Pharmacology Toxicology University of Arkansas for Medical Sciences, Little Rock, AR Lab of Paul L. Prather, Ph.D.

Publications

6. Clayton S, Alexander JS, Solitro G, White L, Villalba S, Winder E, Boudreaux M, Veerareddy P, Dong E, Minagar A, Dao HN, Sorrels D. Self-expanding intestinal expansion sleeves (IES) for short gut syndrome. Ped Surg Internat, 2021 (recently accepted).

5. Vanhoof-Villalba S, Gautier N, and Glasscock E. Pharmacogenetics of KCNQ channel activation in two potassium channelopathy mouse models of epilepsy. Epilepsia 2017, 59: 358-368.

4. Mishra V, Karumuri BK, Gautier N, Liu R, Hutson T, Vanhoof-Villalba S, Vlachos I, Iasemidis L, and Glasscock E. Scn2a deletion improves survival and brain-heart dynamics in the Kcna1-null mouse model of sudden unexpected death in epilepsy (SUDEP). Hum Mol Genet 2017, 26: 2091-2103.

3. Lee S, Holly K, Voziyanov V, Villalba S, Tong R, Grigsby H, Glasscock E, Szele F, Vlachos I, and Murray T. Gradient index microlens implanted in prefrontal cortex of mouse does not affect behavioral test performance over time. PLoS ONE 2016, 11(1): e0146533.

2. Satyanarayanajois S, Villalba S, and Go M. Design, synthesis, and docking studies of peptidomimetics based on HER2-herceptin binding site with potential antiproliferative activity against breast cancer cell lines. Chemical Biology and Drug Design 2009, 74(3):246-57.

1. Martin N, Ruckle M, Vanhoof S, and Prather P. Agonist, antagonist, and inverse agonist characteristics of TIPP (H-Tyr-Tic-Phe-Phe-OH), a selective delta-opioid receptor ligand. The Journal of Pharmacology and Experimental Therapeutics 2002, 301(2):661-71.

&

Current Professional and Service Activities

Institutional

Louisiana State University Shreveport, Shreveport, LA

Graduate Recruitment & Education Committee – Graduate Director (Spring 2022—) IACUC – currently under development Chair Search Committee – Dept. of Biological Sciences (Spring 2021) Faculty Search Committee – Dept. of Biological Sciences (Spring 2021) Innovation Committee (2021-present) Faculty Collaboratory Committee (2021-present) Health Sciences Advisory Committee (2020-present)

Other

Appointment with the Open Educational Resource (OER) for Dual Enrollment Faculty Cohorts from LOUIS: The Louisiana Library Network at the Louisiana Board of Regents (2021-2022)

CBIO 1044: General Biology II Lecture + Lab (Science Majors) – General concepts and principles of ecology, evolution, and biological diversity, for science majors. The course material is presented in a combined lecture and laboratory format.

Professional Memberships

2021-	Faculty for Undergraduate Neuroscience
2021-	Louisiana Academy of Sciences
2019-	Center for Brain Health – LSUHSC-S
2017-2020	American Association for Anatomy
2017-2020	American Association of Clinical Anatomists
2015-	Society for Neuroscience – Shreveport Chapter
2014-2017	American Epilepsy Society

Teaching Interests

- I. Human Anatomy & Physiology
- II. Human Neuroanatomy & Neuroscience
- III. Histology

Research Interests

Dr. Villalba's current research focus is geared towards medical and anatomical educational research and neuroscience research. She collaborates with faculty at both LSU Health Science Center – Shreveport and LSUS. Her neuroscience research, in collaboration with Dr. Kathryn A. Hamilton at LSUHSC-S, focuses on protein and gene expression in olfactory epithelium, concentrating on various stages of development of olfactory sensory neurons. The goal of this project is to further our understanding of neuronal development and plasticity.